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(51) International Patent Classification : C12N 15/52, 9/16, C07K 16/40, C12N 11/14, C12Q 1/42, A61K 38/43	A2	(11) International Publication Number: WO 98/04712 (43) International Publication Date: 5 February 1998 (05.02.98)
(21) International Application Number: PCT/US97/13016 (22) International Filing Date: 24 July 1997 (24.07.97) (30) Priority Data: 08/685,992 25 July 1996 (25.07.96) US (71) Applicant: COLD SPRING HARBOR LABORATORY [US/US]; 1 Bungtown Road, Cold Spring Harbor, NY 11724 (US). (72) Inventors: TONKS, Nicholas; 3 Arrowhead Place, Huntington, NY 11743 (US). FLINT, Andrew, J.; 22711 14th Place West, Bothell, WA 98021 (US). (74) Agents: GRANAHAAN, Patricia et al.; Hamilton, Brook, Smith & Reynolds, P.C., Two Milltia Drive, Lexington, MA 02173 (US).		(81) Designated States: CA, JP, MX, European patent (AT, BE, CH, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE). Published <i>Without international search report and to be republished upon receipt of that report.</i>
(54) Title: SUBSTRATE TRAPPING PROTEIN TYROSINE PHOSPHATASES (57) Abstract Novel protein tyrosine phosphatases in which the invariant aspartate residue is replaced with an alanine residue and which bind to a tyrosine phosphorylated substrate and are catalytically attenuated are described. Also described are methods of identifying tyrosine phosphorylated proteins which complex with the described protein tyrosine phosphatases.		

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SUBSTRATE TRAPPING PROTEIN TYROSINE PHOSPHATASESBackground of the Invention

The protein tyrosine phosphatase (PTP) family of enzymes consists of more than 500 structurally diverse proteins which have in common the highly conserved 250 amino acid PTP catalytic domain, but which display considerable variation in their non-catalytic segments (Charbonneau and Tonks, Annu. Rev. Cell Biol. 8:463-493 (1992); Tonks, Semin. Cell Biol. 4:373-453 (1993)). This structural diversity presumably reflects the diversity of physiological roles of individual PTP family members, which in certain cases have been demonstrated to have specific functions in growth, development and differentiation (Desai et al., Cell 84:599-609 (1996); Kishihara et al., Cell 74:143-156 (1993); Perkins et al., Cell 70:225-236 (1992); Pingel and Thomas, Cell 58:1055-1065 (1989); Schultz et al., Cell 73:1445-1454 (1993)). Although recent studies have also generated considerable information regarding the structure, expression and regulation of PTPs, the nature of the tyrosine phosphorylated substrates through which the PTPs exert their effects remains to be determined. Studies with a limited number of synthetic phosphopeptide substrates have demonstrated some differences in substrate selectivity of different PTPs (Cho et al., Protein Sci. 2: 977-984 (1993); Dechert et al., Eur. J. Biochem. 231:673-681 (1995)), and have indicated preferences for certain amino acid residues at particular positions around the phosphorylated tyrosine residue (Ruzzene et al., Eur. J. Biochem. 211:289-295 (1993); Zhang et al., Biochemistry 33:2285-2290 (1994)). This indicates that PTPs display a certain level of substrate selectivity in vitro, although

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the physiological relevance of the substrates used in these studies is unclear.

Summary of the Invention

As described herein, the substrate specificity of mammalian protein tyrosine phosphatases (PTPs) has been investigated using a novel substrate trapping approach in which mutant or altered forms of the mammalian PTP, also referred to as substrate trapping PTPs, are used to bind (trap) one or more substrates of the PTP. Binding of the substrate trapping PTP with a substrate of the PTP results in the formation of a complex which can be readily observed, and, if desired, isolated, and characterized. The mutant forms of the PTPs have attenuated catalytic activity (lack catalytic activity or have reduced catalytic activity) relative to the wild type PTP but retain the ability to bind tyrosine phosphorylated substrate(s) of the wild type PTP.

The methods of the present invention are specifically exemplified herein with respect to the phosphatases PTP1B and PTP-PEST; however, it is understood that the invention is not limited to these specific PTPs but is applicable to all members of the PTP family. In order to identify potential substrates of PTP1B and PTP-PEST, mutant (i.e., altered or substrate trapping) forms of PTP1B and PTP-PEST were generated which were catalytically attenuated but retained the ability to bind substrates. These mutant PTPs associated in stable complexes with proteins which were identified by immunoblotting as p210 bcr:abl and p130^{cas}, respectively. These associations were observed in lysates from several cell lines and in transfected COS cells, indicating that p210 bcr:abl and p130^{cas} represent major physiologically relevant substrates for PTP1B and PTP-PEST.

These results provide the first demonstration of PTPs having inherently restricted substrate specificity in vivo.

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The methods used to identify p210 bcr:abl and p130^{cas} as specific substrates for PTP1B and PTP-PEST, respectively, are generally applicable to any member of the PTP family, of which approximately 500 members have currently been reported, and can be used to determine the physiological substrates of other members of the PTP family.

One embodiment of the invention relates to novel mutant PTPs in which the invariant aspartate residue is replaced with an amino acid which does not cause significant alteration of the K_m of the enzyme but which results in a reduction in K_{cat} to less than 1 per minute (less than 1 min^{-1}). These PTPs retain the ability to form a complex with, or bind, their tyrosine phosphorylated substrates, but are catalytically attenuated. In one embodiment, the invention relates to the phosphatase PTP1B in which the invariant aspartate residue at position 181 is replaced with alanine (D181A). In another embodiment the invention relates to the phosphatase PTP-PEST in which the invariant aspartate residue at position 199 is replaced with an alanine (D199A). Another embodiment of the invention relates to a PTP-PEST phosphatase in which the cysteine residue at position 231 is replaced with a serine (C231S). The invention also relates to other mutant or substrate trapping PTPs in which the invariant aspartate residue is replaced with or changed to another amino acid residue, such as alanine. The invariant aspartate residue can be identified in other PTPs by aligning the PTP nucleotide sequence with the nucleotide sequence of a PTP for which the location of the invariant aspartate residue is known.

The invention also relates to a method of identifying a tyrosine phosphorylated substrate of a protein tyrosine phosphatase. According to one embodiment of the present invention, a tyrosine phosphorylated protein of interest is combined with one or more PTP(s) in which the invariant aspartate residue is replaced with an amino acid which does

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not cause significant alteration of the K_m of the enzyme but which results in a reduction in K_{cat} to less than 1 per minute (less than 1 min^{-1}), and the presence or absence of a complex between the protein and the PTP(s) is determined.

5 Presence of a complex in the combination indicates that the tyrosine phosphorylated protein is a substrate of the PTP. The PTP DA mutant binds to or complexes with its substrate but does not dephosphorylate it (or does so very slowly), thereby allowing the complex to be observed and, optionally,
10 isolated and identified. In a particular embodiment of the invention, the invariant aspartate is replaced with an alanine residue (a PTP DA mutation or alteration)

In an alternative embodiment of the present invention, a PTP of interest in which the invariant aspartate residue
15 is replaced with an amino acid which does not cause significant alteration of the K_m of the enzyme but which results in a reduction in K_{cat} to less than 1 per minute (less than 1 min^{-1}), is combined with one or more tyrosine phosphorylated proteins, and the presence or absence of a
20 complex between the protein(s) and the PTP is determined. Presence of a complex in the combination indicates that the tyrosine phosphorylated protein is a substrate of the PTP. The PTP DA mutant binds to or complexes with its substrate but does not dephosphorylate it (or does so very slowly),
25 thereby allowing the complex to be observed, and, optionally, isolated and identified. In one embodiment of the invention, the invariant aspartate residue is replaced with an alanine residue (a PTP DA mutation or alteration)

The present invention also relates to a method of
30 identifying a tyrosine phosphorylated substrate of a protein tyrosine phosphatase wherein more than one tyrosine phosphorylated protein of interest is combined with more than one PTP of interest in which the invariant aspartate residue is replaced with an amino acid which does not cause
35 significant alteration of the K_m of the enzyme but which

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results in a reduction in Kcat to less than 1 per minute (less than 1 min^{-1}) (e.g., the invariant aspartate is replaced with an alanine residue). Complexes formed in the combination can be isolated and the component PTP and
5 substrate can be identified.

The invention also pertains to a method of reducing the activity of a tyrosine phosphorylated protein, comprising administering to a mammal a PTP in which the invariant aspartate residue is replaced with an amino acid
10 which does not cause significant alteration of the Km of the enzyme but which results in a reduction in Kcat to less than 1 per minute (less than 1 min^{-1}) (e.g., the invariant aspartate is replaced with an alanine residue) and which forms a complex with the tyrosine phosphorylated protein.
15 The PTP mutant binds to the phosphorylated protein without dephosphorylating it, thereby inhibiting the activity of the protein and reducing its downstream effects.

For example, the invention relates to a method of reducing the transforming effects of oncogenes associated
20 with p130^{cas}, a substrate of PTP-PEST, comprising administering to a mammal wild type PTP-PEST or PTP-PEST in which the invariant aspartate residue is replaced with an alanine residue. Wild type PTP-PEST binds and dephosphorylates p130^{cas}, thereby negatively regulating its
25 downstream effects. DA mutants of PTP-PEST bind but do not dephosphorylate p130^{cas} (or dephosphorylate it at a reduced rate); the substrate is thus tied up in the complex with the substrate trapping form of PTP-PEST and cannot exert its downstream effects. Similarly, the invention relates to a
30 method of reducing the formation of signalling complexes associated with p130^{cas}, particularly those signalling complexes which induce mitogenic pathways, comprising administering to a mammal wild type PTP-PEST or PTP-PEST in which the invariant aspartate residue is replaced with an
35 alanine residue.

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The present invention also relates to assays for identifying agents which alter, e.g., enhance or inhibit, the interaction between a PTP and its phosphorylated substrate. Agents identified by these assays can be
5 agonists (e.g., agents which enhance or increase the activity of the PTP) or antagonists (e.g., agents which inhibit or decrease the activity of the PTP) of PTP activity. The agent may be an endogenous physiological substance or may be a natural or synthetic drug, including
10 small organic molecules.

For example, the tyrosine phosphorylated substrate of a PTP can be identified by the methods described herein. An enzymatic activity assay utilizing the wild type PTP can be carried out in the presence of an agent to be tested, and
15 the resulting amount of enzyme activity can be compared with the amount of enzyme activity in the absence of the agent to be tested. A decrease in the enzymatic activity in the presence of the agent to be tested indicates that the agent inhibits the interaction between the PTP and its substrate.
20 Conversely, an increase in the enzymatic activity in the presence of the agent to be tested indicates that the agent enhances the interaction between the PTP and its substrate.

Alternatively, a competitive binding assay can be carried out utilizing the mutant PTP in the presence of an
25 agent to be tested, and the resulting extent of binding of the mutant PTP to its substrate can be compared with the extent of binding in the absence of the agent to be tested. A decrease in the extent of binding in the presence of the agent to be tested indicates that the agent inhibits the
30 interaction between the PTP and its substrate. Conversely, an increase in the extent of binding in the presence of the agent to be tested indicates that the agent enhances the interaction between the PTP and its substrate.

Thus, the compositions and methods described herein
35 are useful in identifying the tyrosine phosphorylated

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substrates of members of the PTP family of phosphatases, as well as in regulating the activity of identified substrates. The compositions and methods described herein are also useful for identifying tyrosine phosphorylated proteins

5 which are related to a particular disease or disorder, and to methods of screening for modulators which enhance or inhibit the PTP/substrate interaction for use in therapeutic applications.

Brief Description of the Drawings

10 Figures 1A and 1B show a multiple sequence alignment of the catalytic domains of PTPs. In Figure 1A, cytosolic eukaryotic PTPs and domain 1 of RPTPs are combined into one group, domains 2 of RPTPs are in a second group, and the *Yersinia* PTP is in a third. Invariant residues shared among

15 all three groups are shown in red. Invariant and highly conserved residues within a group are shown in blue and green, respectively. Within the *Yersinia* PTP sequence, residue that are either invariant or highly conserved between the cytosolic and RPTP domain sequences are colored

20 blue and green, respectively. The position of residues of PTP1B that interact with the peptide are indicated with a red arrow, and the residue numbering at the bottom of the alignment corresponds to that for PTP1B. Figure 1B is a black and white photocopy of Figure 1A in which the colored

25 areas are indicated with labeled arrows.

Figure 2 shows the Vmax and Km of various PTP1B mutants toward RCML.

Detailed Description of the Invention

The PTP family of enzymes contains a common

30 evolutionarily conserved segment of approximately 250 amino acids known as the PTP catalytic domain. Within this

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conserved domain is a unique signature sequence motif, [I/V]HCXAGXXR[S/T]G, that is invariant among all PTPs. The cysteine residue in this motif is invariant in members of the family and is known to be essential for catalysis. It functions as a nucleophile to attack the phosphate moiety of the incoming substrate. If the cysteine residue is altered by site-directed mutagenesis to serine (CS mutants) or alanine (CA mutants), the resulting PTP is catalytically attenuated but retains the ability to complex with, or bind, its substrate, at least in vitro. These results have been confirmed relative to MKP-1, a member of the PTP family (Sun et al., Cell 75:487-493 (1993)), as well as other PTPs. However, although these CS mutants can in general bind effectively to phosphotyrosyl substrates in vitro, in many cases such complexes cannot be isolated in vivo. Thus, the CS mutants are limited in their applicability and cannot be used to isolate all combinations of PTPs and substrates.

The crystal structures of PTP1B alone (Barford, et al., Science 263:1397-1404 (1994)) and in a complex with a phosphotyrosine-containing peptide (Jia et al., Science 268:1754-1758 (1995)) were recently determined. These structures indicated twenty seven invariant residues (Barford et al., 1994), one of which is an aspartate residue. This aspartate residue is invariant across the catalytic domains of PTP family members. That is, if the amino acid sequences of the PTP family members are aligned, the aspartate residue is present in each PTP at a corresponding location, although the position numbers may be different due to the shifts required to maximize alignment (see the Figure (from Barford et al., Nature Struc. Biol. 2:1043-1053 (1995)) for an alignment of various PTP sequences). Sequences for which the alignment has not yet been published can readily be aligned with other known PTP sequences, e.g., utilizing available computer software such as GENEWORKS.

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Thus, mutant PTPs other than those specifically described herein can readily be made by aligning the amino acid sequence of the PTP catalytic domain with those described herein, identifying the invariant aspartate
5 residue, and changing the residue by site-directed mutagenesis. Although the specific examples of PTP mutants described herein are aspartate to alanine mutants (DA mutants), it is understood that the invention is not limited to changes of aspartate to alanine. The invariant aspartate
10 residue can be changed, e.g., by site-directed mutagenesis, to any amino acid which does not cause significant alteration of the K_m of the enzyme but which results in a reduction in K_{cat} to less than 1 per minute (less than 1 min^{-1}). For example, the invariant aspartate residue can
15 be changed or mutated to an alanine, valine, leucine, isoleucine, proline, phenylalanine, tryptophan, methionine, glycine, serine, threonine, cysteine, tyrosine, asparagine, glutamine, lysine, arginine or histidine.

As described herein, pervanadate-treated cells were
20 used as an abundant source of tyrosine phosphorylated proteins to investigate the substrate specificity of PTP-PEST. PTP-PEST is an 88 kDa cytosolic PTP (Charest *et al.*, Biochem. J. **308**:425-432 (1995); den Hertog *et al.*, Biochem. Biophys. Res. Commun. **184**:1241-1249 (1992); Takekawa *et al.*,
25 Biochem. Biophys. Res. Commun. **189**:1223-1230 (1992); Yang *et al.*, J. Biol. Chem. **268**:6622-6628 (1993); Yang *et al.*, J. Biol. Chem. **268**:17650 (1993)) which is expressed ubiquitously in mammalian tissues (Yi *et al.*, Blood **78**:
2222-2228 (1991)), and which exhibits high specific activity
30 when assayed *in vitro* using artificial tyrosine phosphorylated substrates (Garton and Tonks, EMBO J. **13**:3763-3771 (1994)). It has previously been demonstrated that PTP-PEST is subject to regulation via phosphorylation of Ser39 *in vitro* and *in vivo*. This modification is
35 catalyzed by both protein kinase C (PKC) and protein kinase

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A (PKA), and results in reduced enzyme activity as a consequence of an increase in the K_m of the dephosphorylation reaction (Garton and Tonks, EMBO J. 13:3763-3771 (1994)). It appears likely that further
5 regulatory mechanisms exist for PTP-PEST, since this enzyme would be expected to exert a considerable negative influence on the tyrosine phosphorylation state of cytosolic substrates of tyrosine kinases. One possibility is that this influence could be limited by the substrate specificity
10 of PTP-PEST.

The substrate specificity of PTP1B was investigated utilizing the same methods outlined for PTP-PEST, with the exception that the cells were not treated with pervanadate. A combination of in vitro dephosphorylation and substrate
15 trapping experiments were used to study the substrate interactions of PTP1B and PTP-PEST. The substrate trapping methods outlined herein are generally applicable to any PTP by virtue of the shared invariant aspartate residue, and should therefore prove useful in delineating the substrate
20 preference of other PTP family members. In particular, the use of mutant, catalytically impaired PTPs to trap, and thereby isolate, potential substrates will greatly facilitate the identification of physiologically important substrates for individual PTPs, leading to improved
25 understanding of the roles of these enzymes in regulation of cellular processes.

One embodiment of the invention relates to novel PTPs in which the invariant aspartate residue is replaced with an amino acid which does not cause significant alteration of
30 the K_m of the enzyme but which results in a reduction in K_{cat} to less than 1 per minute (less than 1 min^{-1}). These PTPs retain the ability to form a complex with, or bind, their tyrosine phosphorylated substrates but are catalytically attenuated. As defined herein, "attenuated"
35 activity is intended to mean that the phosphatase retains a

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similar K_m to that of the wild type phosphatase but has a V_{max} which is reduced by a factor of at least 10^4 relative to the wild type enzyme. This includes catalytic activity which is either reduced or abolished relative to the wild type PTP. For example, the invariant aspartate residue can be changed or mutated to an alanine, valine, leucine, isoleucine, proline, phenylalanine, tryptophan, methionine, glycine, serine, threonine, cysteine, tyrosine, asparagine, glutamine, lysine, arginine or histidine.

10 The novel PTPs described herein, in which the invariant aspartate residue is replaced with an amino acid which does not cause significant alteration of the K_m of the enzyme but which results in a reduction in K_{cat} to less than 1 per minute (less than 1 min^{-1}), can also comprise other
15 mutations, particularly those which assist in stabilizing the PTP/substrate complex. For example, a mutation of the [serine/threonine] residue in the signature motif to an alanine residue changes the rate-determining step of the dephosphorylation reaction from the formation of the
20 transition state to the break down of the transition state, thereby stabilizing the PTP/substrate complex. Such mutations may be valuably combined with the replacement of the invariant aspartate residue, particularly assisting in stabilizing the complex and facilitating the observation and
25 isolation of the complex.

PTPs suitable for use in the invention include any PTP which has an invariant aspartate residue in a corresponding position. As defined herein, a phosphatase is a member of the PTP family if it contains the signature motif
30 [I/V]HCXAGXXR[S/T]G. Dual specificity PTPs, i.e., PTPs which dephosphorylate both phosphorylated tyrosine and phosphorylated serine or threonine, are also suitable for use in the invention. Appropriate PTPs include, but are not limited to, PTP1B, PTP-PEST, PTP $_{\gamma}$, MKP-1, DEP-1, PTP $_{\mu}$,
35 PTPX1, PTPX10 and PTPH1.

In one embodiment, the invention relates to the phosphatase PTP1B in which the aspartate residue at position 181 is replaced with alanine (D181A). In another embodiment the invention relates to the phosphatase PTP-PEST in which
5 the invariant aspartate residue at position 199 is replaced with an alanine (D199A). Another embodiment of the invention relates to a PTP-PEST phosphatase in which the cysteine residue at position 231 is replaced with a serine (C231S).

10 The invention also relates to a method of identifying a tyrosine phosphorylated protein which is a substrate of a particular protein tyrosine phosphatase. According to one embodiment of the present invention, a tyrosine
15 phosphorylated protein of interest is combined with at least one PTP in which the invariant aspartate residue is replaced with an amino acid which does not cause significant alteration of the K_m of the enzyme but which results in a reduction in K_{cat} to less than 1 per minute (less than 1 min^{-1}) (e.g., an alanine residue), and the presence or
20 absence of a complex between the protein and the PTP is determined. Presence of a complex in the combination indicates that the tyrosine phosphorylated protein is a substrate of the PTP. The PTP DA mutant (substrate trapping mutant) binds to or complexes with its substrate but does
25 not dephosphorylate it (or does so very slowly), thereby allowing the complex to be isolated and identified.

The phosphorylated protein/PTP complex may be isolated by conventional isolation techniques as described in U.S. Patent No. 5,352,660 to Pawson, including salting out,
30 chromatography, electrophoresis, gel filtration, fractionation, absorption, polyacrylamide gel electrophoresis, agglutination, or combinations thereof. Furthermore, to facilitate the determination of the presence of the protein/PTP complex, antibodies against the PTP or
35 the phosphorylated protein can be used, as well as labelled

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PTPs and/or labelled phosphorylated substrates. The PTP or phosphorylated protein can be labelled with various enzymes, fluorescent materials, luminescent materials and radioactive materials. Examples of suitable enzymes include, but are not limited to, horseradish peroxidase, biotin, alkaline phosphatase, β -galactosidase and acetylcholinesterase. Examples of suitable fluorescent materials include, but are not limited to, umbelliferone, fluorescein, fluorescein isothiocyanate, rhodamine, dichlorotriazinylamine fluorescein, dansyl chloride and phycoerythrin. Appropriate luminescent materials include luminol, and suitable radioactive material include radioactive phosphorous ^{32}P , iodine I^{125} , I^{131} or tritium.

Alternatively, the invention pertains to a method of identifying a tyrosine phosphorylated protein which is a substrate of a PTP, comprising combining a PTP of interest in which the invariant aspartate residue is replaced with an amino acid which does not cause significant alteration of the K_m of the enzyme but which results in a reduction in K_{cat} to less than 1 per minute (less than 1 min^{-1}) (e.g., an alanine residue), with at least one tyrosine phosphorylated protein, thereby producing a combination; and determining the presence or absence of a complex in the combination, wherein presence of a complex in the combination between a tyrosine phosphorylated protein and the PTP indicates that the tyrosine phosphorylated protein is a substrate of the PTP.

The substrate trapping PTPs of the present invention can also be used in place of wild type PTPs to screen phosphotyrosyl peptide libraries for peptides which bind to the PTP as described in Songyang *et al.* (*Nature* 373:536-539 (1995); *Cell* 72:767-778 (1993)). Peptides identified from such peptide libraries can then be assessed to determine whether tyrosine phosphorylated proteins containing these peptides exist in nature.

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Any tyrosine phosphorylated protein is suitable as a potential substrate in the present invention. Tyrosine phosphorylated proteins are well known in the art. Specific examples of appropriate substrates include, without
5 limitation, p130^{cas}, the EGF receptor, p210 bcr:abl, MAP kinase and the insulin receptor. Of particular interest are tyrosine phosphorylated proteins which have been implicated in a mammalian disease or disorder.

The invention also pertains to a method of reducing
10 the activity of a tyrosine phosphorylated protein, comprising administering to a mammal a PTP in which the invariant aspartate residue is replaced with an amino acid which does not cause significant alteration of the Km of the enzyme but which results in a reduction in Kcat to less than
15 1 per minute (less than 1 min⁻¹) (e.g., an alanine residue) and which forms a complex with the tyrosine phosphorylated protein. The PTP DA mutant binds to the phosphorylated protein without dephosphorylating it (or causing dephosphorylation at a greatly reduced rate), thereby
20 inhibiting the activity of the protein and reducing its downstream effects. As used herein, "reducing" includes both reduction and complete abolishment, e.g., of one or more activities or functions of the phosphorylated protein.

For example, the invention relates to a method of
25 reducing the transforming effects of oncogenes associated with p130^{cas}, a substrate of PTP-PEST, comprising administering to a mammal wild type PTP-PEST or PTP-PEST in which the invariant aspartate residue is replaced with an alanine residue. Wild type PTP-PEST binds and
30 dephosphorylates p130^{cas}, thereby negatively regulating its downstream effects. DA mutants of PTP-PEST bind but do not dephosphorylate p130^{cas} (or do so at a greatly reduced rate); the substrate is thus tied up in the complex with the substrate trapping form of PTP-PEST and cannot exert its
35 downstream effects. Similarly, the invention relates to a

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method of reducing the formation of signalling complexes associated with $p130^{cas}$, particularly those signalling complexes which induce mitogenic pathways, comprising administering to a mammal wild type PTP-PEST or PTP-PEST in which the invariant aspartate residue is replaced with an alanine residue. The PTP binds to and/or dephosphorylates $p130^{cas}$, thereby negatively regulating the downstream effects of $p130^{cas}$ and reducing the formation of signalling complexes associated with $p130^{cas}$.

The substrate trapping mutant PTPs of the present invention can be used in virtually any application in place of, or in addition to, a corresponding wild type PTP. The advantages of such a utility lie in the ability of the mutant PTP to mimic the function of the wild type enzyme, e.g., to decrease the activity of its tyrosine phosphorylated substrate, without inducing the harmful cytotoxic effects commonly observed with administration or overexpression of the wild type PTP. Thus, the invention also pertains to a method of reducing the cytotoxic effects associated with administration or overexpression of wild type PTPs. For example, CS mutants of MKP-1 have been shown to have the same functional effect as wild type MKP-1 without induction of potentially harmful side effects. Thus, PTPs described herein, in which the invariant aspartate residue is replaced with an amino acid which does not cause significant alteration of the K_m of the enzyme but which results in a reduction in K_{cat} to less than 1 per minute (less than 1 min^{-1}) (e.g., an alanine residue), can be used in many applications in place of the corresponding wild type enzyme. As used herein, a "corresponding" enzyme is one which is the same as the mutant PTP (e.g., PTP-PEST and PTP-PEST D199A) or one which is different from the mutant PTP but recognizes the same substrate as the mutant PTP.

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The mutant PTPs described herein can also be used therapeutically to reduce the activity of a tyrosine phosphorylated protein, such as by a gene therapy method in which the mutant PTP described herein, or a functional

5 portion thereof which retains the ability to bind to its tyrosine phosphorylated substrate, is introduced into a subject and in whom the mutant PTP is expressed. The mutant PTP replaces, either partially or totally, the wild type enzyme which is normally produced or competes with the wild

10 type PTP for binding to the substrate. For example, a specific tyrosine phosphorylated protein can be identified which is implicated in a particular disease or disorder (such as a protein tyrosine kinase). At least one PTP which acts to dephosphorylate the selected tyrosine phosphorylated

15 protein of the present invention can be identified by the methods described herein. The wild type or mutant form of the PTP can be administered to a subject in need of treatment in order to tie up or bind the tyrosine phosphorylated substrate, thereby inhibiting or reducing the

20 function of the phosphorylated protein. In a preferred embodiment, the mutant PTP is administered in place of the wild type enzyme in order to reduce the cytotoxic effects associated with overexpression of the wild type enzyme. Procedures for gene therapy are known in the art (see U.S.

25 Patent No. 5,399,346 to Anderson et al.) and can be modified by methods known in the art to appropriately express the specific mutant and wild type PTPs of the present invention.

The present invention also relates to assays for identifying agents which alter, e.g., enhance or inhibit,

30 the interaction between a PTP and its phosphorylated substrate. Agents identified by these assays can be agonists (e.g., agents which enhance or increase the activity of the PTP) or antagonists (e.g., agents which inhibit or decrease the activity of the PTP) of PTP

35 activity. The agent may be an endogenous physiological

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substance or may be a natural or synthetic drug, including small organic molecules.

For example, the tyrosine phosphorylated substrate of a PTP can be identified by the methods described herein. An enzymatic activity assay utilizing the wild type PTP can be carried out in the presence of an agent to be tested, and the resulting amount of enzyme activity can be compared with the amount of enzyme activity in the absence of the agent to be tested. Enzymatic activity assays are known in the art; for example, assays of PTP activity using a tyrosine phosphorylated ³²P-labelled substrate are described in Flint *et al.* (*EMBO J.* 12:1937-1946 (1993)). A decrease in the enzymatic activity in the presence of the agent to be tested indicates that the agent inhibits the interaction between the PTP and its substrate. Conversely, an increase in the enzymatic activity in the presence of the agent to be tested indicates that the agent enhances the interaction between the PTP and its substrate.

Alternatively, a competitive binding assay can be carried out utilizing the mutant PTP in the presence of an agent to be tested, and the resulting extent of binding of the mutant PTP to its substrate can be compared with the extent of binding in the absence of the agent to be tested. Competitive binding assays are known in the art; for example, U.S. Patent No. 5,352,660 to Pawson describes methods suitable for use in this invention. A decrease in the extent of binding in the presence of the agent to be tested indicates that the agent inhibits the interaction between the PTP and its substrate. Conversely, an increase in the extent of binding in the presence of the agent to be tested indicates that the agent enhances the interaction between the PTP and its substrate.

According to the present invention, tyrosine phosphorylated peptides identified with mutant PTPs from peptide libraries by the methods of Songyang *et al.* (*Nature*

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373:536-539 (1995); Cell 72:767-778 (1993)) can be used herein in place of the complete tyrosine phosphorylated protein in competitive binding assays.

The present invention also pertains to pharmaceutical compositions comprising a PTP in which the invariant aspartate residue is replaced with an amino acid which does not cause significant alteration of the K_m of the enzyme but which results in a reduction in K_{cat} to less than 1 per minute (less than 1 min^{-1}) (e.g., an alanine residue). For instance, the PTP of the present invention can be formulated with a physiologically acceptable medium to prepare a pharmaceutical composition. The particular physiological medium may include, but is not limited to, water, buffered saline, polyols (e.g., glycerol, propylene glycol, liquid polyethylene glycol) and dextrose solutions. The optimum concentration of the active ingredient(s) in the chosen medium can be determined empirically, according to procedures well known to medicinal chemists, and will depend on the ultimate pharmaceutical formulation desired. Methods of introduction of exogenous PTPs at the site of treatment include, but are not limited to, intradermal, intramuscular, intraperitoneal, intravenous, subcutaneous, oral and intranasal. Other suitable methods of introduction can also include rechargeable or biodegradable devices and slow release polymeric devices. The pharmaceutical compositions of this invention can also be administered as part of a combinatorial therapy with other agents.

The following Examples are offered for the purpose of illustrating the present invention and are not to be construed to limit the scope of this invention. The teachings of all references cited herein are hereby incorporated by reference in their entirety.

EXAMPLESMaterials and Methods

The following is a description of the materials and methods used in the work described herein.

5 Generation, Expression and Purification of Mutant PTP
Proteins

Point mutations within the catalytic domains of PTP-
PEST (D199A, C231S) and PTP1B (D181A, C215S) were introduced
by site-directed mutagenesis using the Muta-GeneTM in vitro
10 mutagenesis kit (Bio-Rad, Richmond, CA). Regions containing
the required point mutation were then exchanged with the
wild type sequences within appropriate expression vectors,
and the replaced mutant regions were sequenced in their
entirety to verify the absence of additional mutations.

15 Full length PTP-PEST proteins (wild type and mutant
proteins, containing either Asp199 to Ala or Cys231 to Ser
mutations) and the wild type PTP-PEST catalytic domain
(amino acids 1-305) were expressed in Sf9 cells using
recombinant baculovirus (BaculoGoldTM, Pharmingen, San
20 Diego, CA), and purified as described in Garton and Tonks
(EMBO J. 13:3763-3771 (1994)). Truncated forms of wild type
and mutant PTP-PEST proteins, comprising amino acid residues
1-305 of PTP-PEST were also expressed in E. coli as GST
fusion proteins following subcloning of PTP-PEST DNA in-
25 frame downstream of GST in pGEX vectors (Pharmacia Biotech
Inc., Uppsala, Sweden). Twenty-five ml of E. coli
transformed with the appropriate vector were grown to log
phase (OD₆₀₀ approximately 0.5). Fusion protein expression
was then induced by addition of 0.2 mM isopropyl-1-thio-β-D-
30 galactopyranoside, and the cells were grown for 2-4 hours at
30°C. Cells were harvested by centrifugation, incubated
with 50 mg/ml lysozyme in 3 ml buffer containing 50 mM Tris-
HCl, pH 7.4, 5mM EDTA, 1 mM PMSF, 1 mM benzamidine, 5 mg/ml
leupeptin, 5 mg/ml aprotinin, 0.1 % Triton X-100 and 150 mM

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NaCl, then lysed by sonication (3 x 10s). Following removal of insoluble material by centrifugation (20 minutes at 300,000 x g), fusion proteins were isolated by incubation for 30 min at 4°C with 100 ml glutathione-Sepharose beads (Pharmacia Biotech Inc., Uppsala, Sweden), and the beads were then collected by centrifugation and washed three times with buffer A (20 mM Tris-HCl, pH 7.4, 1 mM EDTA, 1 mM benzamidine, 1 mg/ml leupeptin, 1 mg/ml aprotinin, 10 % glycerol, 1 % Triton X-100 and 100 mM NaCl). This procedure yielded essentially homogeneous fusion protein at a concentration of 1 mg protein/ml glutathione-Sepharose beads. PTP1B proteins (wild type and mutant forms) comprising amino acids 1-321 were expressed in *E.coli* and purified to homogeneity as described in Barford *et al.* (*J. Mol. Biol.* 239:726-730 (1994)).

Cell Culture, Transfection, Preparation of Lysates and Fractionation

HeLa and COS cells were grown in Dulbecco's modified Eagle's medium (DMEM), containing 5% fetal bovine serum (FBS); Wi38, C2C12 and MvLu cells were grown in DMEM containing 10% FBS; 293 cells were grown in DMEM containing 10% calf serum; MCF10A cells were grown in 50% DMEM, 50% Ham's F-12 containing 5% horse serum, 20 ng/ml epidermal growth factor, 10 mg/ml insulin, 0.5 mg/ml hydrocortisone and 0.25 mg/ml fungizone. All media also contained penicillin and streptomycin at 100 U/ml and 100 mg/ml, respectively, and all cells were grown at 37°C. Calcium phosphate-mediated transfection was used to introduce cDNA encoding wild type and mutant PTP-PEST proteins into COS cells. These were encoded by PTP-PEST cDNA subcloned into the plasmid PMT2, from which expression was driven by an adenovirus major late promoter; 20 mg DNA was used for transfection of each 10 cm plate of cells. The level of expression of PTP-PEST constructs was similar in all cases.

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Prior to cell lysis, 70-90% confluent cultures of cells were treated for 30 minutes with 0.1 mM pervanadate (20 ml of a fresh solution containing 50 mM sodium metavanadate (NaVO_3) and 50 mM H_2O_2 were added to 10 ml medium). Treatment of cells with H_2O_2 and vanadate leads to a synergistic increase in phosphotyrosine levels, presumably due to inhibition of intracellular PTPs by vanadate. The synergism between H_2O_2 and vanadate has previously been suggested to result from improved accumulation of the resultant oxidized vanadate (pervanadate) within the cells when compared to vanadate itself (Heffetz *et al.*, *J. Biol. Chem.* 265:2896-2902 (1990)). It is important to note that during the preparation of cell lysates, dilution occurs such that the inhibitory effect of vanadate on PTP action is lost. Pervanadate treatment resulted in the appearance of at least 50 prominent phosphotyrosine protein bands in all cell types, whereas untreated cells contained virtually undetectable levels of phosphotyrosine (data not shown).

Cells were lysed in Buffer A containing 5 mM iodoacetic acid, which was included in order to inhibit irreversibly cellular PTPs. Following incubation at 4°C for 30 minutes, 10 mM DTT was added to inactivate any unreacted iodoacetic acid. Insoluble material was then removed by centrifugation for 20 minutes at 300,000 x g. The resultant lysates were stable with regard to their phosphotyrosine content during long term (several months) storage at -70°C and during prolonged (at least 20 hours) incubation at 4°C, in the absence of exogenous added PTPs.

Pervanadate-treated HeLa cell lysate was fractionated by anion exchange chromatography using a Mono Q FPLC column (Pharmacia). The sample (50 mg total protein at 3 mg/ml in buffer A) was diluted in three volumes of buffer B (20 mM tris-HCl, pH 7.4, 1 mM EDTA, 1 mM benzamidine, 1 mg/ml leupeptin, 1 mg/ml aprotinin and 0.1% Triton X-100) prior to

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loading. Proteins were eluted at a flow rate of 1 ml/min with a linear gradient of 0-0.5 M NaCl in buffer B over 20 fractions (1 ml fraction volume), followed by a second gradient of 0.5-1.0 M NaCl in buffer B over 5 fractions.

- 5 Phosphotyrosine-containing proteins were detected within fractions 7-21 according to anti-phosphotyrosine immunoblotting. The same procedures were followed for PTP1B, with the exception that the cells were not treated with pervanadate.

10 Dephosphorylation Reactions

- Lysates of pervanadate-treated HeLa cells (1-2 mg protein/ml) containing tyrosine phosphorylated proteins were incubated on ice in the absence or presence of purified active PTPs at a concentration of 2 nM. Dephosphorylation
15 was terminated by the removal of aliquots (30 mg protein) into SDS-PAGE sample buffer, and the extent of dephosphorylation was determined by immunoblotting using the monoclonal antibody G104. Assays of PTP activity using tyrosine phosphorylated ³²P-labelled RCM-lysozyme as
20 substrate were performed as described in Flint *et al.* (*EMBO J.* 12:1937-1946 (1993)).

Antibodies and Immunoblotting

- The PTP-PEST monoclonal antibody AG25 was raised against baculovirus-expressed purified full-length PTP-PEST.
25 The anti-phosphotyrosine monoclonal antibody G104 was generated using as antigen phosphotyrosine, alanine and glycine, in a 1:1:1 ratio, polymerized in the presence of keyhole limpet hemocyanin with 1-ethyl-3-(3'-dimethylaminopropyl)carbodiimide, a method originally
30 described in Kamps and Sefton (*Oncogene* 2:305-315 (1988)). p130^{cas} monoclonal antibody was from Transduction Laboratories (Lexington, Ky). Monoclonal antibody FG6 against PTP1B was provided by Dr David Hill (Calbiochem

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Oncogene Research Products, Cambridge, MA). Visualization of proteins by immunoblotting was achieved by enhanced chemiluminescence (ECL) using HRP-conjugated secondary antibodies (Amersham Life Science Inc., Arlington Heights, 5 Il) and the SuperSignalTM CL-HRP substrate system (Pierce, Rockford, Il).

Immunoprecipitation and Substrate Trapping

Immunoprecipitation of PTP-PEST from transfected COS cells was performed following covalent coupling of 10 monoclonal antibody AG25 to protein A-Sepharose beads (Pharmacia Biotech Inc., Uppsala, Sweden) using the chemical cross-linking agent dimethyl pimelimidate (Schneider et al., J. Biol. Chem. 257:10766-10769 (1982)). Antibody was first bound to protein A-Sepharose at a concentration of 1 mg/ml 15 bead volume, and unbound material was then removed by three washes with 0.2 M sodium borate, pH 9. Covalent coupling was achieved by incubation at room temperature for 30 minutes in the presence of 20 mM dimethyl pimelimidate in 0.2 M sodium borate, pH 9. The beads were then incubated 20 for 1 hour with an excess of 0.2 M ethanolamine, pH 8, to block any unreacted cross-linker, and washed three times with PBS prior to storage at 4°C. Ten ml of AG25 beads were used to precipitate transfected PTP-PEST from lysates containing approximately 0.375 mg protein.

25 Substrate trapping was performed using various PTP affinity matrices. The full-length PTP-PEST matrix utilized covalent coupled AG25-protein A-Sepharose beads to which purified baculovirus-expressed PTP-PEST protein was bound. Aliquots (10ml) of AG25 beads were incubated for 2 hours at 30 4°C in 100 ml buffer A in the presence of 5 mg of purified PTP-PEST (wild type or mutant forms); unbound PTP-PEST was then removed by washing three times with 1 ml buffer A. The resultant PTP-PEST-AG25-protein A-Sepharose beads contained approximately 2 mg of PTP-PEST per 10 ml aliquot. Substrate

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trapping was also carried out with glutathione-Sepharose beads bound to bacterially-expressed GST fusion proteins containing the catalytic domain of PTP-PEST.

PTP1B was also used in substrate trapping experiments.

- 5 In this case, the monoclonal antibody FG6 was precoupled to protein A-Sepharose in the absence of cross-linker (2 mg antibody/10 ml beads), then purified PTP1B proteins were added in excess and incubated at 4°C for 2 hours. Following removal of unbound PTP1B, 10 ml beads contained
- 10 approximately 2 mg PTP1B.

- Pervanadate-treated cell lysates, or column fractions, were used as a source of phosphotyrosine-containing proteins for substrate trapping experiments. In general, lysates containing 0.25-0.5 mg protein in 0.5 ml buffer A (including
- 15 5 mM iodoacetic acid, 10 mM DTT) were incubated at 4°C for 2 hours in the presence of 10 ml of affinity matrix containing approximately 2 mg of the appropriate PTP protein. Unbound proteins were then removed from the samples by washing three times with 1 ml buffer A, and bound material was collected
- 20 by addition of 50 ml SDS-PAGE sample buffer followed by heating at 95°C for 5 minutes; proteins bound to the beads were then analyzed by SDS-PAGE followed by immunoblotting.

Results

- The following details the results of the work
- 25 described herein carried out as described above.

PTP1B and p210 bcr:abl

- Chronic myelogenous leukemia (CML) is a clonal disorder of the haematopoietic stem cell that is characterized by the Philadelphia chromosome, in which the
- 30 c-Abl proto-oncogene on chromosome 9, encoding a PTK, becomes linked to the bcr gene on chromosome 22. This results in the generation of a bcr:abl fusion protein, p210 bcr:abl, in which the PTK activity is enhanced relative to

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that of c-Abl. Current data indicates that this cytogenetic abnormality is the primary and sole causative event in CML. Expression of p210 bcr:abl produces abnormal patterns of tyrosine phosphorylation that result in the aberrant
5 maturation of the haematopoietic stem cell that is characteristic of CML.

Expression of PTP1B mRNA and protein is enhanced as a consequence of p210 bcr:abl expression in Rat1, Mo7 and BaF3 cells. Changes in PTP1B activity, which were commensurate
10 with the change in enzyme protein, were also observed. These changes are specific for PTP1B and are not seen in closely related homologue (65% identity) TC-PTP or in other tested PTKs, including SHP-1, SHP-2 and PTP-PEST. The increase in expression of PTP1B was also observed in Ph+ B-
15 lymphoid cells derived from a CML patient relative to Ph-cells from the same patient.

The changes in PTP1B levels were induced specifically by p210 bcr:abl and were not seen in cells expressing other PTKs including v-abl, v-src or other oncoproteins such as
20 myc. The PTK activity of p210 bcr:abl was essential for the increase in expression of PTP1B, since expression of an inactive lysine to arginine mutant form of p210 bcr:abl in Rat1 cells did not alter PTP1B levels. The increase in PTP1B levels is a rapid response to induction of p210
25 bcr:abl. When BaF3 cells expressing a temperature-sensitive mutant form of p210 bcr:abl were shifted to the permissive temperature for the PTK, PTP1B levels were observed to increase within 12-24 hours coincident with the appearance of the active form of the PTK. These data indicate that the
30 alteration PTP1B levels is a relatively rapid response to the appearance of p210 bcr:abl, rather than a long-term adaptive response of the cells.

In transient cotransfection experiments in COS cells, PTP1B dephosphorylates p210 bcr:abl but not v-abl. When the
35 PTP1B D181A mutant was expressed as a GST fusion protein,

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purified and incubated with lysates of Mo7-p210 cells (which overexpress p210 bcr:abl), a complex of the mutant PTP and p210 bcr:abl was isolated. In contrast, tyrosine phosphorylated c-abl, which was also present in the lysates, did not bind to the mutant PTP. The interaction between PTP1B D181A and p210 bcr:abl was blocked by vanadate, suggesting that the interaction involved the active site of the PTP.

Following transient coexpression in COS cells, PTP1B D181A formed a complex with p210 bcr:abl. Preliminary data indicate that the Y177F mutant form of p210 bcr:abl did not interact with PTP1B D181A, suggesting that this tyrosine residue is a component of the binding site in the PTK. This tyrosine residue in p210 bcr:abl is phosphorylated in vivo and has been demonstrated to serve as a docking site for GRB2. Direct interaction of the pTyr in p210 bcr:abl and the SH2 domain of GRB2 is essential for the transforming activity of the PTK. Interaction of PTP1B D181A with p210 bcr:abl interferes with the association of the PTK with GRB2. Taken together, these data suggest that p210 bcr:abl is a physiological substrate of PTP1B and that PTP1B may function as an antagonist of the oncoprotein PTK in vivo. The Vmax, Km and Kcat of 37 kDa PTP1B mutants toward RCML are shown in Figure 2.

25 PTP1B and the EGF Receptor

Expression of PTP1B D181A in COS cells leads to enhanced phosphorylation of tyrosyl residues in a 180 kDa protein and in proteins of 120 and 70 kDa. When a GST-PTP1B D181A fusion protein is expressed in COS cells and precipitated on Glutathione-Sepharose, the 180 kDa, and smaller quantities of p120 and p70, were coprecipitated. The p180 protein was identified as the epidermal growth factor (EGF) receptor by immunoblotting. The identity of

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the p120 and p70 proteins is unclear; however, the latter is not src, p62 or paxillin.

Expression of PTP1B D181A in COS cells induces tyrosine phosphorylation of the EGF receptor in the absence
5 of its ligand, EGF, indicating that the mutant PTP is exerting its effects in the intact cell and not post-lysis. The equivalent D199A PTP-PEST mutant does not interact with the EGF receptor, indicating the specificity of this substrate interaction.

10 Autophosphorylation of the EGF receptor is required for the interaction with PTP1B D181A. Mutants of the receptor that are either kinase-dead or in which the autophosphorylation sites have been deleted do not interact with PTP1B D181A. In v-src-expressing cells, a plethora of
15 tyrosine phosphorylated proteins were observed, but phosphorylation of the EGF receptor was not detected. Under these conditions, PTP1B D181A bound predominantly to a 70 kDa tyrosine phosphorylated protein.

As a result of this work, it appears that PTP1B can
20 modulate EGF-induced signalling pathways, perhaps including the pathways of many diseases, including breast cancer.

Preferential Dephosphorylation of a 130 kDa Phosphotyrosine-Containing Protein by PTP-PEST

In order to investigate the substrate specificity of
25 PTP-PEST in vitro, aliquots of pervanadate-treated HeLa cell lysates were incubated on ice, yielding 50-100 distinct phosphotyrosine-containing proteins as judged by immunoblotting of the cell lysate using the monoclonal anti-phosphotyrosine antibody G104. Purified full-length PTP-
30 PEST (expressed in Sf9 cells using recombinant baculovirus), PTP-PEST catalytic domain, or PTP1B catalytic domain (37 kDa form) was then added to the lysate, and aliquots were removed at various time points for analysis by SDS-PAGE followed by anti-phosphotyrosine immunoblotting.

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Surprisingly, a prominent 130 kDa phosphotyrosine band (p130) was selectively dephosphorylated by PTP-PEST within 10 minutes, whereas the intensity of all the other bands was essentially unchanged even after 60 minutes of incubation with PTP-PEST. Long incubations with higher concentrations of PTP-PEST (greater than 100-fold) resulted in the complete removal of all phosphotyrosine bands from the lysate. However, under all conditions tested, p130 was found to be dephosphorylated more rapidly than all other bands present.

The selective dephosphorylation of p130 by PTP-PEST was also observed using a truncated form of the phosphatase (amino acid residues 1-305) which essentially contains only the catalytic domain of the enzyme. This result suggests that the striking substrate preference displayed by PTP-PEST in this analysis is an inherent property of the phosphatase catalytic domain, whereas the C-terminal 500 amino acid residues have little discernible effect on the substrate specificity of the enzyme.

The specificity of the interaction between PTP-PEST and p130 was addressed using the catalytic domain of PTP1B (amino acid residues 1-321) in dephosphorylation reactions. When added at a similar molar concentration to that used for PTP-PEST, PTP1B was found to dephosphorylate fully and rapidly (within 15 minutes) most of the phosphotyrosine-containing proteins present in the pervanadate-treated HeLa lysate. In addition, the time course of dephosphorylation of p130 was not significantly more rapid than that of the other phosphotyrosine bands dephosphorylated by PTP1B. It should be noted, however, that these in vitro dephosphorylation results are not truly illustrative of the substrate specificity of PTP1B in vivo for several reasons. First, only the isolated catalytic subunit was used in this particular experiment. Furthermore, in vivo substrate specificity may be quite different due to the intracellular distribution of both the PTP and potential substrates. That

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is, in vitro dephosphorylation experiemnts may utilize substrates which the PTP is capable of dephosphorylating but which it would not have access to in vivo. The phenomenon of differing substrate specificity depending upon different physiologic contexts is illustrated by a comparison of this data with the in vivo PTP1B work described above, wherein PTP1B showed specificity for only three proteins.

Identification of Phosphotyrosine-Containing p130 Protein as p130^{cas} by Substrate Trapping

10 Pervanadate-treated HeLa cell lysate was fractionated by anion exchange chromatography and aliquots of the fractions were analyzed by SDS-PAGE followed by immunoblotting with anti-phosphotyrosine or anti-p130^{cas} antibodies. Aliquots of all samples analyzed were then
15 incubated with an affinity matrix containing a substrate trapping PTP-PEST mutant, comprising full length PTP-PEST in which Asp199 is changed to alanine (D199A), bound to covalently coupled protein A-Sepharose/antibody (AG25) beads. Proteins associated with PTP-PEST were then analyzed
20 by SDS-PAGE followed by immunoblotting with anti-phosphotyrosine or anti-p130^{cas} antibodies.

Anti-phosphotyrosine immunoblotting of the column fractions showed that the p130 phosphotyrosine band eluted as a single peak in fractions 11-14 (approx. 0.3 M NaCl).
25 In view of the abundance of tyrosine phosphorylated p130 in HeLa lysates, it appeared likely that p130 represents a previously identified phosphotyrosine-containing 130 kDa protein. Several potential candidates were identified in the literature, including the focal adhesion kinase p125^{FAK}, ras-GAP, gp130 and p130^{cas}. Of these candidates,
30 p130^{cas} has been identified as a particularly prominent phosphotyrosine band in a wide variety of systems, including v-crk (Mayer and Hanafusa, Proc. Natl. Acad. Sci. USA 87: 2638-2642 (1990); Mayer et al., Nature 332:272-275 (1988))

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and src (Kanner et al., Proc. Natl. Acad. Sci. USA 87:3328-3332 (1990); Reynolds et al., Mol. Cell. Biol. 9: 3951-3958 (1989)) transformed fibroblasts, integrin-mediated cell adhesion (Nojima et al., J. Biol. Chem. 270:15398-15402 (1995); Petch et al., J. Cell Science 108:1371-1379 (1995); Vuori and Ruoslahti, J. Biol. Chem. 270:22259-22262 (1995)) and PDGF stimulated 3T3 cells (Rankin and Rozengurt, J. Biol. Chem. 269:704-710 (1994)).

Therefore, the possibility that the p130
10 phosphotyrosine band corresponds to p130^{cas} was tested by immunoblotting the Mono Q fractions using an antibody to p130^{cas}. The 130 kDa band corresponding to p130^{cas} eluted in the same fractions as the p130 tyrosine phosphorylated band, and displayed a similar apparent molecular weight,
15 suggesting that they might represent the same protein. Furthermore, p130^{cas} immunoprecipitated from these fractions was found to be phosphorylated on tyrosyl residues.

A mutant form of PTP-PEST (D199A) was generated by
20 site-directed mutagenesis, and the mutant enzyme was purified following expression using recombinant baculovirus. When assayed using tyrosine phosphorylated RCM-Lysozyme as substrate, the purified mutant enzyme exhibited a specific activity which was approximately 10,000 fold lower than
25 that of the wild type enzyme (Garton and Tonks, unpublished data). This purified protein was bound to an affinity matrix comprised of an anti-PTP-PEST monoclonal antibody (AG25) covalently coupled to Protein A-Sepharose beads, then incubated with each of the Mono Q fractions. After 45
30 minutes of incubation, proteins associating with the mutant PTP-PEST were collected by centrifugation, the beads were washed, and SDS-PAGE sample buffer was added. Associated proteins were then analyzed by immunoblotting using the monoclonal anti-phosphotyrosine antibody G104.

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The mutant PTP-PEST protein was found to associate with a single phosphotyrosine-containing protein, the molecular weight (130 kDa) and Mono Q elution position (fractions 11-14) of which coincided with those of p130^{cas}. Immunoblotting of the PTP-PEST-associated proteins using the p130^{cas} antibody demonstrated that the 130 kDa tyrosine phosphorylated protein trapped by the mutant PTP-PEST is indeed p130^{cas}. These data further support the hypothesis that p130^{cas} is a potential physiologically relevant substrate for PTP-PEST.

Determination of Structural Features of PTP-PEST Involved in Specific Interaction with Tyrosine Phosphorylated p130^{cas}

The interaction between p130^{cas} and PTP-PEST was investigated further in substrate trapping experiments using various purified mutant forms of PTP-PEST to precipitate proteins from pervanadate-treated HeLa lysates. Several affinity matrices were incubated with pervanadate-treated HeLa cell lysate, and proteins associated with the beads were analyzed by SDS-PAGE followed by immunoblotting with anti-phosphotyrosine or anti-p130^{cas} antibodies.

The wild type full-length phosphatase was found to be incapable of stable association with tyrosine phosphorylated p130^{cas}, whereas both the PTP-PEST (D199A) mutant protein and a mutant lacking the active site cysteine residue (C231S) specifically precipitated p130^{cas} from the lysate. The inability of the wild type phosphatase to precipitate tyrosine phosphorylated p130^{cas} presumably reflects the transient nature of the normal interaction between PTP-PEST and tyrosine phosphorylated p130^{cas}, which is likely to be concluded as soon as p130^{cas} is dephosphorylated by PTP-PEST.

Since the C-terminal 500 amino acids of PTP-PEST contain several proline-rich regions which resemble src homology-3 (SH3) domain binding sequences, it appeared

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plausible that the specificity of the interaction between PTP-PEST and p130^{cas} might depend to some extent on association of these segments with the SH3 domain of p130^{cas}. The possible contribution of the C-terminal
5 segment of PTP-PEST in the observed specific interaction of PTP-PEST with p130^{cas} was therefore addressed in further substrate trapping experiments using GST fusion proteins containing the catalytic domain of PTP-PEST alone, in both wild type and mutant (D199A) forms. The mutant catalytic
10 domain of PTP-PEST fused to GST was found to precipitate the p130^{cas} phosphotyrosine band specifically, whereas both the wild type fusion protein and GST alone failed to precipitate p130^{cas}. The specific interaction between PTP-PEST and p130^{cas} observed in these experiments therefore appears to
15 be an intrinsic property of the catalytic domain of PTP-PEST, emulating the observed preference of the active PTP-PEST catalytic domain for dephosphorylation of p130^{cas} in vitro.

Specificity of Interaction Between Mutant PTP-PEST and Tyrosine Phosphorylated p130^{cas}

In view of the relative abundance of tyrosine phosphorylated p130^{cas} in the pervanadate-treated HeLa cell lysate, the possibility that the observed selective binding of PTP-PEST inactive mutant proteins to p130^{cas} was
25 substrate-directed (reflecting the abundance of this potential substrate relative to the other phosphotyrosine-containing proteins present in the lysate) rather than enzyme-directed (reflecting a genuine substrate preference of PTP-PEST) was considered; this possibility was addressed
30 in two ways. First, inactive mutant forms of the catalytic domain of PTP1B were used to trap potential substrates for this enzyme from the pervanadate-treated HeLa lysates. Again it was found that the wild type phosphatase was incapable of stable interaction with any phosphotyrosine-

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containing protein, whereas mutant variants of the PTP1B phosphatase domain (comprising Cys or Asp mutations analogous to those described above for PTP-PEST) associated with many tyrosine phosphorylated proteins. This was especially apparent for the aspartic acid mutant of PTP1B (D181A), which appeared to precipitate essentially all phosphotyrosine-containing proteins from the lysate with similar efficacy. These data emphasize the specific nature of the interaction between PTP-PEST and p130^{cas}, which appears to be a property peculiar to the PTP-PEST catalytic domain, rather than a feature shared by all PTP catalytic domains.

The specificity of the interaction between PTP-PEST and p130^{cas} was addressed further following pervanadate-treatment of several different cell lines (Wi38, 293, COS, MCF10A, C2C12, MvLu), yielding a different array of tyrosine phosphorylated proteins in each case; the resultant lysates were analyzed by SDS-PAGE followed by anti-phosphotyrosine immunoblotting. Aliquots were incubated with PTP-PEST (D199A) affinity matrix or control matrix, and tyrosine phosphorylated proteins associating with PTP-PEST were analyzed by SDS-PAGE and immunoblotting with anti-phosphotyrosine or anti-p130^{cas} antibodies as described above.

In each case, the D199A mutant PTP-PEST protein precipitated a single broad phosphotyrosine band with an apparent molecular weight between 120 and 150 kDa in different cell lines, whereas the affinity matrix alone failed to precipitate any phosphotyrosine-containing protein. Immunoblotting of the precipitates with a p130^{cas} antibody revealed that the protein precipitated from all cell lysates corresponded to p130^{cas}; the observed molecular weight variation between different cell lines presumably reflects either species differences in the molecular weight of p130^{cas} or expression of different

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alternatively spliced forms (Sakai *et al.*, *EMBO J.* 13:3748-3756 (1994)).

The relative abundance of tyrosine phosphorylated p130^{cas} in the PTP-PEST precipitates appeared to correlate approximately with the abundance of p130^{cas} protein in the lysates (data not shown). Surprisingly, regardless of the abundance of tyrosine phosphorylated p130^{cas} in the lysates, p130^{cas} was invariably the only phosphotyrosine-containing protein in the precipitates, even in 293 cell lysates which contained very little p130^{cas} protein but which displayed a wide variety of other abundantly tyrosine phosphorylated proteins. Similarly, when lysates of pervanadate-treated 293 cells (containing tyrosine phosphorylated p130^{cas} in amounts which are undetectable by anti-phosphotyrosine immunoblotting of the lysate) were incubated with active PTP-PEST, no visible dephosphorylation of any phosphotyrosine band occurred (Garton and Tonks, unpublished data). These results indicate that the affinity of PTP-PEST for p130^{cas} is substantially greater than for any other substrate present, and further emphasizes the remarkable substrate selectivity of PTP-PEST for p130^{cas}.

Vanadate Inhibition of Tyrosine Phosphorylated p130^{cas} Association with Mutant PTP-PEST

A consistent observation of this work was that, in contrast to the inactive mutant PTP-PEST, the wild type enzyme failed to associate in a stable complex with tyrosine phosphorylated p130^{cas}, suggesting that the observed association is active site-directed. In order to investigate this possibility, mutant PTP-PEST (D199A) was incubated with the PTP inhibitor vanadate at various concentrations prior to addition of pervanadate-treated HeLa cell lysate. The extent of association of p130^{cas} with PTP-PEST was then analyzed. PTP-PEST affinity matrix, comprising full length PTP-PEST (D199A) bound to covalently

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coupled protein A-Sepharose/antibody (AG25) beads, was incubated for 10 minutes on ice in the presence of varying concentrations of sodium orthovanadate. The samples were then incubated with aliquots of pervanadate-treated HeLa
5 cell lysate; associated proteins were analyzed by SDS-PAGE and immunoblotting with anti-phosphotyrosine or anti-p130^{cas} antibodies. The activity of wild type PTP-PEST was also determined under the same conditions, using tyrosine phosphorylated ³²P-labelled RCM-lysozyme as substrate.

10 The association was found to be potently disrupted by vanadate, with a concentration-dependence similar to that of vanadate inhibition of wild type PTP-PEST, and complete disruption being observed at 10 mM vanadate. Since PTP inhibition by vanadate presumably results from a direct
15 interaction of vanadate with the active site cysteine residue of the enzyme (Denu *et al.*, *Proc. Natl. Acad. Sci. USA* 93:2493-2498 (1996)), this result supports the hypothesis that the stable association of mutant PTP-PEST with tyrosine phosphorylated p130^{cas} is mediated by direct
20 interactions between active site residues within PTP-PEST, in particular the active site cysteine residue, and phosphotyrosine moieties within p130^{cas}.

Association of Endogenous p130^{cas} with Transfected Mutant PTP-PEST in COS Cells

25 The work described above strongly suggests that p130^{cas} represents a potential physiologically significant substrate for PTP-PEST. In order to assess whether PTP-PEST interacts with p130^{cas} in intact cells, COS cells were transfected with plasmids encoding wild type or mutant forms
30 of PTP-PEST (D199A or C215S). The cells were treated with pervanadate 30 minutes prior to lysis, PTP-PEST proteins were immunoprecipitated, and associated tyrosine phosphorylated proteins were analyzed by anti-phosphotyrosine immunoblotting of the resultant

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precipitates. Lysates were also incubated with covalently coupled protein A-Sepharose/anti-PTP-PEST (AG25) beads and associated proteins were analyzed by SDS-PAGE and immunoblotting with anti-phosphotyrosine antibody.

5 Under these conditions, the phosphotyrosine-containing band corresponding to p130^{cas} was again unique in its ability to associate with the C231S PTP-PEST protein, indicating that p130^{cas} can be specifically selected by PTP-PEST as a substrate in an intracellular context in the
10 presence of a large number of alternative possible substrates. Neither the wild type nor the D199A form of PTP-PEST was capable of a stable interaction with tyrosine phosphorylated p130^{cas} in pervanadate-treated COS cells.

The binding of both wild type and D199A PTP-PEST to
15 tyrosine phosphorylated p130^{cas} under these conditions is most likely prohibited by the presence of pervanadate bound to the active site cysteine residue of PTP-PEST (Denu et al., Proc. Natl. Acad. Sci. USA 93:2493-2498 (1996)), which effectively excludes the binding of phosphotyrosine residues
20 of p130^{cas}. The ability of the C231S mutant PTP-PEST to associate in a stable complex with p130^{cas} in the presence of pervanadate suggests that this mutant protein is largely unaffected by pervanadate, indicating that the normal mode of inhibition of PTPs by vanadate ions depends critically on
25 direct interactions between vanadate and the thiolate anion of the PTP active-site cysteine residue. These observations therefore lend further support to the existence of an exclusive interaction between PTP-PEST and p130^{cas} which appears to be entirely active site-directed, and therefore
30 reflects the genuine, inherent, highly restricted substrate preference of PTP-PEST for p130^{cas}.

Results described herein implicate p130^{cas} as a physiologically relevant substrate for PTP-PEST. Furthermore, the observed stringency and exclusivity of the
35 interaction between PTP-PEST and p130^{cas} in a wide variety

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of cell lines suggest that p130^{cas} may be a unique high affinity substrate for PTP-PEST, although the possibility that other significant PTP-PEST substrates may exist cannot be excluded at present. In particular, it is unclear whether pervanadate-treated cells display a complete spectrum of all possible tyrosine phosphorylated proteins; in fact this appears unlikely since pervanadate treatment presumably results only in an increase in tyrosine phosphorylation of proteins which are to some extent constitutively phosphorylated, but which are normally rapidly dephosphorylated, within the cell. Potential substrates lacking from pervanadate-treated cells therefore presumably include substrates of protein tyrosine kinases (PTKs) which are normally present in an inactive state, such as ligand-stimulated receptor PTKs, and the recently described calcium regulated kinase PYK2 (Lev *et al.*, Nature 376: 737-745 (1995)). Regardless of these considerations, the ability of PTP-PEST to select p130^{cas} exclusively as a substrate from lysates of several different cell lines, containing a combined total of at least one hundred different potential substrates (many of which presumably contain multiple sites of phosphorylation), clearly demonstrates that the substrate specificity of PTP-PEST is highly restricted.

Many intracellular PTPs are limited in their substrate availability due to strict confinement within a particular subcellular location; examples include PTP1B, which is localized to the cytoplasmic face of the endoplasmic reticulum (Frangioni *et al.*, Cell 68:545-560 (1992)), and TCPTP which is either nuclear (Tillmann *et al.*, Mol. Cell. Biol. 14:3030-3040 (1994)) or localized to the endoplasmic reticulum, depending upon which alternative spliced form is expressed (Lorenzen *et al.*, J. Cell Biol. 131:631-643 (1995)). Alternatively, certain PTPs appear to be highly regulated, requiring activation before appreciable activity

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- can be demonstrated. For example, the SH2 domain-containing PTPs, SHP1 and SHP2, display relatively low activity in vitro, but can be considerably activated by several mechanisms including C-terminal truncation (Zhao et al., J. Biol. Chem. 268:2816-2820 (1993)), addition of certain phospholipids (Zhao et al., Proc. Natl. Acad. Sci. USA 90:4251-4255 (1993)), or SH2 domain-mediated binding of appropriate phosphotyrosine-containing peptides (Lechleider et al., J. Biol. Chem. 268:21478-21481 (1993)).
- 10 However, PTP-PEST exhibits high specific activity in vitro (35,000 U/mg), and is a predominantly (90-95%) soluble PTP within cells (Garton and Tonks, unpublished data); in principle, therefore, it may act potently on any substrate accessible to the cytoplasm. This accessibility may partly
- 15 underlie the necessity for PTP-PEST to possess an inherently constrained substrate specificity. The demonstration that mutant PTP-PEST is capable of exclusively associating with p130^{cas} in an intracellular context in the presence of many other tyrosine phosphorylated proteins, is an indication
- 20 that the narrow substrate specificity of the enzyme may result in PTP-PEST having a negligible influence on the phosphorylation state of the majority of tyrosine phosphorylated proteins within the cell, even though those substrates are largely accessible to PTP-PEST.
- 25 The role of p130^{cas} in cellular transformation by the v-crk and v-src oncogenes is unclear, although there is a general correlation between the level of tyrosine phosphorylation of p130^{cas} and the degree of transformation in cells expressing different forms of crk or src (Kanner et al., EMBO J. 10:1689-1698 (1991); Mayer and Hanafusa, J. Virol. 64:3581-3589 (1990)). Furthermore, enhanced tyrosine phosphorylation of p130^{cas} has also been observed in cells transformed by c-Ha-ras and by ornithine decarboxylase overexpression (Auvinen et al., Mol. Cell. Biol. 15:6513-6525 (1995)). Expression of antisense cDNA encoding
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p130^{cas} in these cells results in a partial reversion of the transformed phenotype. These observations suggest that aberrant tyrosine phosphorylation of p130^{cas} is a common feature of cells transformed by several disparate mechanisms and that p130^{cas} may be required for full manifestation of the transformed state. Dephosphorylation of p130^{cas} by PTP-PEST is therefore a potentially important regulatory mechanism for counteracting the transforming effects of various oncogenes.

10 Tyrosine phosphorylation of p130^{cas} has been observed in fibroblasts following integrin-mediated cell adhesion to extracellular matrix proteins (Nojima et al., J. Biol. Chem. 270:15398-15402 (1995); Petch et al., J. Cell Science 108:1371-1379 (1995); Vuori and Ruoslahti, J. Biol. Chem. 15 270:22259-22262 (1995)). Under these conditions, using an antibody (4F4) that predominantly recognizes tyrosine phosphorylated p130^{cas} (Kanner et al., EMBO J. 10:1689-1698 (1991); Petch et al., J. Cell Science 108:1371-1379 (1995)) it was shown that phosphorylated p130^{cas} is localized to 20 focal adhesions (Petch et al., J. Cell Science 108:1371-1379 (1995)), whereas fractionation studies have demonstrated that the normal cellular location of the majority of non-phosphorylated p130^{cas} is the cytosol (Sakai et al., EMBO J. 13:3748-3756 (1994)). Furthermore, in crk-transformed 25 fibroblasts, tyrosine phosphorylated p130^{cas} is detected only in insoluble fractions (Sakai et al., EMBO J. 13:3748-3756 (1994)), suggesting that both cell adhesion- and transformation-mediated phosphorylation of p130^{cas} is associated with redistribution of the protein from the 30 cytosol to focal adhesions.

It is plausible that the redistribution of tyrosine phosphorylated p130^{cas} may be driven by its association with FAK, which is constitutively associated with focal adhesions due to its C-terminal focal adhesion targeting 35 domain (Hildebrand et al., J. Cell Biol. 123:993-1005

(1993); Schaller *et al.*, Proc. Natl. Acad. Sci. USA **89**:5192-5196 (1992)). The sequestration of tyrosine phosphorylated p130^{cas} in focal adhesions both in transformed cells, and following integrin-mediated cell adhesion, strongly suggests a role for p130^{cas} in signalling events in this region of the cell. One consequence of the redistribution of tyrosine phosphorylated p130^{cas} is likely to be that, in addition to localizing p130^{cas} to a region of the cell containing abundant protein tyrosine kinase activity, the phosphorylated protein will be relatively inaccessible to the cytosolic phosphatase PTP-PEST. This raises the possibility that the role of PTP-PEST in dephosphorylating p130^{cas} may be to prevent inappropriate tyrosine phosphorylation of the cytosolic pool of p130^{cas}, thus preventing formation of signalling complexes assembled around tyrosine phosphorylated p130^{cas} in inappropriate cellular locations.

Several mitogenic factors potently stimulate tyrosine phosphorylation of p130^{cas}. These include agents acting through heterotrimeric G protein-coupled receptors such as lysophosphatidic acid (Seufferlein and Rozengurt, J. Biol. Chem. **269**:9345-9351 (1994)), bradykinin (Leeb-Lundberg *et al.*, J. Biol. Chem. **269**: 24328-24344 (1994)), and bombesin (Zachary *et al.*, J. Biol. Chem. **267**:19031-19034 (1992)), as well as growth factors that activate receptor tyrosine kinases, namely PDGF (Rankin and Rozengurt, J. Biol. Chem. **269**:704-710 (1994)), EGF and NGF (Ribon and Saltiel, J. Biol. Chem. **271**:7375-7380 (1996)). These observations suggest roles for p130^{cas} in regulation of mitogenic signalling pathways, presumably involving assembly of signalling complexes based on tyrosine phosphorylated p130^{cas}. The identities of the proteins involved in these complexes are not established, but are likely to include SH2 domain-containing adaptor proteins such as crk (Ribon and Saltiel, J. Biol. Chem. **271**:7375-7380 (1996)), and its

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associated proteins (Feller et al., Oncogene 10:1465-1473 (1995); Hasegawa et al., Mol. Cell. Biol. 16:1770-1776 (1996); Knudsen et al., J. Biol. Chem. 269:32781-32787 (1994); Matsuda et al., Mol. Cell. Biol. 14: 5495-5500 (1994); Tanaka et al., Proc. Natl. Acad. Sci. USA 91:3443-3447 (1994)). Therefore tyrosine phosphorylation and dephosphorylation of p130^{cas} potentially plays a central role in regulating the formation of such complexes, thereby influencing downstream events in mitogenic signalling.

10 Equivalents

Those skilled in the art will recognize, or be able to ascertain, using no more than routine experimentation, many equivalents to the specific embodiments of the invention described herein. Such equivalents are intended to be encompassed by the following claims:

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CLAIMS

We claim:

1. A protein tyrosine phosphatase wherein the invariant aspartate residue is replaced with an amino acid (e.g. alanine) which:
 - a) does not cause significant alteration of the K_m of the enzyme but which results in a reduction in K_{cat} to less than 1 per minute; or
 - b) is selected from the group consisting of:
alanine, valine, leucine, isoleucine, proline, phenylalanine, tryptophan, methionine, glycine, serine, threonine, cysteine, tyrosine, glutamine, lysine, arginine and histidine, and wherein the phosphatase binds to a tyrosine phosphorylated substrate and is catalytically attenuated.
2. A protein tyrosine phosphatase according to claim 1(a) which is selected from the group consisting of: PTP1B (and wherein for example the invariant aspartate residue is located at position 181), PTP-PEST (and wherein for example the invariant aspartate residue is located at position 199), PTP τ , MKP-1, DEP-1, PTP μ , PTPX1, PTPX10 and PTPH1.
3. A protein tyrosine phosphatase according to claim 1(b) which is a PTP-PEST phosphatase in which the amino acid at position 231 is replaced with a serine residue.
4. A method of identifying a tyrosine phosphorylated protein which is a substrate of a protein tyrosine phosphatase, comprising the steps of:
 - a) combining at least one tyrosine phosphorylated protein with at least one protein tyrosine

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- phosphatase in which the invariant aspartate residue is replaced with an amino acid (e.g. alanine) which does not cause significant alteration of the K_m of the enzyme but which results in a reduction in K_{cat} to less than 1 per minute, under conditions appropriate for formation of a complex between the tyrosine phosphorylated protein and the protein tyrosine phosphatase, thereby producing a combination; and
- 5 b) determining the presence of absence of a complex in the combination wherein the presence of a complex in the combination indicates that the tyrosine phosphorylated protein is a substrate of the protein tyrosine phosphatase with which it forms a complex.
- 10 5. A method according to claim 4, wherein the protein tyrosine phosphatase is as defined in any one of claims 1-3.
- 20 6. A method according to claim 4 or claim 5 wherein the tyrosine phosphorylated protein is selected from the group consisting of: p130^{cas}, the EGF receptor, p210 bcr:abl, MAP kinase and the insulin receptor.
- 25 7. A kit for identifying a tyrosine phosphorylated protein substrate of a protein tyrosine phosphatase comprising:
- 30 a) at least one protein tyrosine phosphatase in which the invariant aspartate residue is replaced with an amino acid (e.g. alanine) which does not cause significant alteration of the K_m of the enzyme but which results in a reduction in K_{cat} to less than 1 per minute; and

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- b) ancillary reagents suitable for use in detecting the presence or absence of a complex between the protein tyrosine phosphatase and a tyrosine phosphorylated protein,
5 wherein for example the protein tyrosine phosphatase is as defined in any one of claims 1-3.
8. A method of identifying an agent which alters the interaction between a protein tyrosine phosphatase and a tyrosine phosphatase, comprising the steps of:
- 10 a) identifying a tyrosine phosphorylated protein which is a substrate of a protein tyrosine phosphatase;
- b) combining the tyrosine phosphorylated protein and the protein tyrosine phosphatase and an agent to
15 be tested under conditions suitable for interaction between the tyrosine phosphorylated protein and the protein tyrosine phosphatase, thereby forming a combination;
- c) determining the amount of enzymatic activity in
20 the combination; and
- d) comparing the amount of enzymatic activity determined in (c) with the amount of enzymatic activity in the absence of the agent to be tested, under conditions suitable for interaction
25 between the tyrosine phosphorylated protein and the protein tyrosine phosphatase,
wherein a difference in the enzymatic activity indicates that the agent alters the interaction between the protein tyrosine phosphatase and the
30 tyrosine phosphorylated protein.
9. A method of identifying an agent which alters the interaction between a protein tyrosine phosphatase and a tyrosine phosphorylated protein which is a substrate

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of the protein tyrosine phosphatase, comprising the steps of:

- 5 a) identifying a tyrosine phosphorylated protein which is a substrate of a protein tyrosine phosphatase;
- 10 b) combining the tyrosine phosphorylated protein, a protein tyrosine phosphatase in which the invariant aspartate residue is replaced with an amino acid (e.g. alanine) which does not cause significant alteration of the K_m of the enzyme but which results in a reduction in K_{cat} to less than 1 per minute, and an agent to be tested, under conditions suitable for interaction between the tyrosine phosphorylated protein and the protein tyrosine phosphatase, thereby forming a combination;
- 15 c) determining the extent of binding between the tyrosine phosphorylated protein and the protein tyrosine phosphatase in the combination; and
- 20 d) comparing the extent of binding determined in (c) with the extent of binding in the absence of the agent to be tested, under conditions suitable for interaction between the tyrosine phosphorylated protein and the protein tyrosine phosphatase,
- 25 wherein a difference in the extent of binding indicates that the agent alters the interaction between the protein tyrosine phosphatase and the tyrosine phosphatase and the tyrosine phosphorylated protein.

- 30 10. A method according to claim 8 or claim 9 wherein if the amount of enzymatic activity or the extent of binding, respectively, is:
 - a) greater in the presence of the agent to be tested than in the absence of the agent, then the agent

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enhances the interaction between the protein tyrosine phosphatase and the tyrosine phosphorylated protein; or

- 5 b) less in the presence of the agent to be tested than in the absence of the agent, then the agent inhibits the interaction between the protein tyrosine phosphatase and the tyrosine phosphorylated protein.

11. A protein tyrosine phosphatase (e.g. as defined in any one of claims 1-3) for use in therapy, prophylaxis or diagnosis, for example in:

- 10 a) the treatment of conditions in which a reduction in the activity of a tyrosine phosphorylated protein is indicated; and/or
- 15 b) reducing the activity of a tyrosine phosphorylated protein; and/or
- 20 c) a method of reducing the activity of a tyrosine phosphorylated protein, comprising administering to a mammal (e.g. a human) a protein tyrosine phosphatase in which the invariant aspartate residue is replaced with an amino acid which does not cause significant alteration of the Km of the enzyme but which results in a reduction in Kcat to less than 1 per minute, whereby formation of
- 25 the complex reduces the activity of the tyrosine phosphorylated protein; and/or
- 30 d) the treatment of conditions in which a reduction in the transforming effects of oncogenes associated with p130^{cas} phosphorylation is indicated; and/or
- e) reducing the transforming effects of oncogenes associated with p130^{cas} phosphorylation; and/or
- f) a method of reducing the transforming effects of oncogenes associated with p130^{cas} phosphorylation

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- comprising administering to a mammal (e.g. a human) a protein tyrosine phosphatase which is PTP-PEST or PTP-PEST in which the invariant aspartate residue is replaced with an amino acid which does not cause significant alteration of the K_m of the enzyme but which results in a reduction in K_{cat} to less than 1 per minute, whereby the PTP binds to and/or dephosphorylates $p130^{cas}$ and reducing the transforming effects of oncogenes associated with $p130^{cas}$ phosphorylation; and/or
- g) the treatment, therapy, diagnosis or prophylaxis of cancer (for example cancers associated with $p130^{cas}$ phosphorylation); and/or
- h) the treatment of conditions associated with oncogenic activity (e.g. with v-crk, v-src and/or c-Ha-ras activity); and/or
- i) the treatment of conditions in which a reduction in the formation of signalling complexes associated with $p130^{cas}$ is indicated; and/or
- j) reducing the formation of signalling complexes associated with $p130^{cas}$; and/or
- k) a method of reducing the formation of signalling complexes associated with $p130^{cas}$ comprising administering to a mammal (e.g. a human) a protein tyrosine phosphatase which is PTP-PEST or PTP-PEST in which the invariant aspartate residue is replaced with an amino acid which does not cause significant alteration of the K_m of the enzyme but which results in a reduction in K_{cat} to less than 1 per minute, whereby the PTP binds to and/or dephosphorylates $p130^{cas}$ and reducing the formation of signalling complexes associated with $p130^{cas}$, and/or

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- 1) the prevention of the induction of mitogenic pathways;
 - m) the treatment of conditions in which the prevention of the induction of mitogenic pathways is indicated;
 - 5 n) the treatment of conditions in which a reduction in the cytotoxic effects associated with protein tyrosine phosphatase administration or over-expression is indicated; and/or
 - 10 o) reducing the cytotoxic effects associated with protein tyrosine phosphatase administration or over-expression; and/or
 - p) a method for reducing the cytotoxic effects associated with protein tyrosine phosphatase administration or over-expression comprising administering to a mammal (e.g. a human) a protein tyrosine phosphatase in which the invariant aspartate residue is replaced with an amino acid which does not cause significant alteration of the K_m of the enzyme but which results in a reduction in K_{cat} to less than 1 per minute, in place of a corresponding wild type protein tyrosine phosphatase; and/or
 - 20 q) in a method according to any one of claims 13-20.
- 25 12. Use of a protein tyrosine phosphatase for the manufacture of a medicament for treatment, prophylaxis or diagnosis (for example for use in the treatments defined in claim 11).
13. A method of reducing the activity of a tyrosine phosphorylated protein, comprising administering to a mammal (e.g. a human) a protein tyrosine phosphatase in which the invariant aspartate residue is replaced with an amino acid which does not cause significant
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alteration of the K_m of the enzyme but which results in a reduction in K_{cat} to less than 1 per minute, whereby formation of the complex reduces the activity of the tyrosine phosphorylated protein.

- 5 14. A method according to claim 13, wherein the tyrosine phosphorylated protein is selected from the group consisting of: p130^{cas}, the EGF receptor, p210 bcr:abl, MAP kinase and the insulin receptor.
- 10 15. a method according to claim 13, wherein the protein tyrosine phosphatase is selected from the group consisting of: PTP1B, PTP-PEST, PTP τ , MKP-1, DEP-1, PTP τ , PTPX1, PTPX10 AND PTPH1.
- 15 16. A method of reducing the transforming effects of oncogenes associated with p130^{cas} phosphorylation comprising of administering to a mammal (e.g. a human) a protein tyrosine phosphatase which is PTP-PEST or PTP-PEST in which the invariant aspartate residue is replaced with an amino acid which does not cause significant alteration of the K_m of the enzyme but
- 20 which results in a reduction in K_{cat} to less than 1 per minute, whereby the PTP binds to and/or dephosphorylates p 130^{cas}, thereby negatively regulating the downstream effects of p130^{cas} and reducing the transforming effects of oncogenes associated with
- 25 p130^{cas} phosphorylation.
17. A method according to claim 16, wherein the oncogene is selected from the group consisting of: v-crk, v-src and c-Ha-ras.

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18. A method of reducing formation of signalling complexes associated with p130^{cas} comprising administering to a mammal (e.g. a human) a protein tyrosine phosphatase which is PTP-PEST OR PTP-PEST in which the invariant aspartate residue is replaced with an amino acid which does not cause significant alteration of the Km of the enzyme but which results in a reduction in Kcat to less than 1 per minute, whereby the PTP binds to and/or dephosphorylates p130^{cas}, thereby negatively regulating the downstream effects of p130^{cas} and reducing the formation of signalling complexes associated with p130^{cas}.
19. A method according to claim 18, which prevents the induction of mitogenic pathways.
20. A method of reducing cytotoxic effects associated with protein tyrosine phosphatase administration or over expression, comprising administering to a mammal (e.g. a human) a protein tyrosine phosphatase in which the invariant aspartate residue is replaced with an amino acid which does not cause significant alteration of the Km of the enzyme but which results in a reduction in Kcat to less than 1 per minute, in place of a corresponding wild type protein tyrosine phosphatase.

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Hum_PTP1B	70	DYINASLI	80	90	100	110	120	130
Hum_TCPTP		DYINASLV		KMEEAQRSYILTQGPLPNTCCHFWMVWE			QKSRGVVMLNRVM	
Hum_PTP_xi_D1		DGKLTdyINANYV		DIEEAQRSYILTQGPLPNTCCHFWMVWQ			QKTKAVVMLNRIV	
Hum_PTP_zela_D1		DGKLTdyINANYV		DGYN RPKAYIAAGGPLKSTAEDEFWRMIWE			HNVEVIVMITNLV	
Hum_PTP_gamma_D		DSKHSdyINANYV		DGYN RPKAYIAAGGPLKSTAEDEFWRMIWE			HNVEVIVMITNLV	
Dros_PTP99A_D1		KKNL dyINANFI		DGYN KAKAYIATQGPLKSTAEDEFWRMIWE			QNTGIIIVMITNLV	
Hum_LCA_D1		PGS dyINANYI		DGYQ KGHAFIGTQGPLPETMGDFWRMVWE			QRTATVVMVTRLE	
Hum_PTP_mu_D1		TNS dyINANYI		DGYR KQNAVYIATQGPLPETMGDFWRMVWE			ENTASIIIMVTNLV	
Hum_PTP_alpha_D1		PDS dyINASFI		NGYH RPNHYIATQGPMEQETIYDFWRMVWH			QNTATIVMVTNLK	
Hum_PTP_opsilon_D		PCS dyINASFI		NGYQ EKNKFIAAAGPKQETVND FWRMVWE			QKSATIVMVTNLK	
Mouso_CD45_D1		AGS TyINASFI		DGFK EPRKYIAAGPRDETVD FWRMVWE			QKATIVMVTNCE	
Hum_SH.PTP2		EPV SDYINANIIMPEFETKCNNSK		PKKSYIATQGCCLQNTVND FWRMVWQ			ENSRVIVMTTKEV	
Hum_SH.PTP1		IPG SDYINANYIKNQLL		GPDE NAKTYIASQGCLEATVND FWRMVWQ			ENSRVIVMTTREV	
Hum_PTP_bola		PCS dyINASFI		PGNN FRREYIVTQGPLPGTKDD FWRMVWE			QNVHNIVMTQCV	
Dros_PTP10D		EGS dyINANYV		PGHN SPREFIVTQGPLHSTRDD FWRMVWE			SNSRAIVMVTNCF	
Hum_SAP.1		PGS dyINASFM		PGLW SPQEFIVTQGPLPQTVGD FWRMVWE			QQSHTLVMLTNCM	
Ral_PTP_STEP		DPL SsyINANYI		RGYNGEEKVYIATQGPVSTVVD FWRMVWQ			ERTPIIIVMITNIE	
Dros_PTP69A_D1		QTT dyINANFV		IGYK ERKFKICAQGMESTIDD FWRMVWE			QHLEIIVILTNE	
Hum_MEG2		TQT dyINASFM		DGYK QKNAYIGTQGPLENTYRD FWRMVWE			QKVLIVMTTREF	
Hum_PTP_PEST		QDS dyINANFI		KGvy GPkAYVATQGPLANTVID FWRMVWE			YNVVIVMACREF	
Hum_PTPH1		INA SyVNMFI		PAAN LVNKYIATQGPLPHTCAQ FWRMVWD			QKLSIIVMLTTLT	
Dici_PTP1		EGS dyINANYI		DGAY PKQFICTQGPLPNTIAD FWRMVWE			NRCRIIVMLSRES	
Fiss_yeast_pyp1		EL dyINASFI		KTETSNYIACQGSISRSISDFWRMVWDNVENIGTIVMLGSLF			QKLSIIVMLTTLT	
Fiss_yeast_pyp2		S dyINASHI		DYGNKKYIACQAPKPGTLLDFWEMVWHNSGIVIMLTNLX			HNVEVIIMITNLV	
Hum_PTP_xi_D2		DGKLTdyINANYV		DGYN RPKAYIAAGGPLKSTAEDEFWRMIWE			HNSTIIIMLTCLR	
Hum_LCA_D2		EGS dyINASFL		DGYR QOKAYIATQGPLAEDEFWRMLWE			WKSCSIIVMLTELE	
Hum_PTP_alpha_D2		ENT dyVNASFI		DGYR QKDSYIASQGPLLHTIED FWRMIWE			WKSHTIIVMLTEVQ	
Hum_PTP_opsilon_D2		EYT dyINASFI		DGYR QKDYFIATQGPLAHTVED FWRMIWE			YHCTSVVMLNDVD	
Hum_PTP_mu_D2		SS NyINAAALM		DSYK QPSAFIVTQHPLPNTVKDFWRLVLD			RKVKVIIVMLTELV	
Mouse_CD45_D2		SEETSKyINASFV		MSYV KPEMMIAAAGPLKETIGDFWQMIFQ			QSVTTLVMISEIG	
Dros_PTP69A_D2		ENS TyINASFI		EGDY NSETFIAAGDPFENTIGDFWRMISE			HNAQLVVMIPDQ	
Hum_PTP_zela_D2		GT dyINASFI		MGYD QSNFIIITQHPLLHTIKDFWRMIWD			HNAQIIIVMLPDNQ	
Hum_PTP_gamma_D2		KGT dyINASFI		MGYD RSNEFIITQHPLPHTTKDFWRMIWD			HNAQTYVVLSSLD	
Dros_PTP99A_D2		GEDGSDyINASWL		HGFR RLRFDFIVLGHFMAHTIKDFWQMVD			NRTPLVLAIVASSS	
Yarsinia_PTP		NyIOVG		NRTIACQYFLOSLESHERMLAE				
PTP1Bseq.no.								

PTP1B66

Hum_PTP1B	1	DFPCRVAKL	10	PNKKN	20	RNRYRDVSPFDH	30	SRIKLHQE	40	50	60	DN
Hum_TCPTP		DYPHRVAKFP		ENRN		RNRYRDVSPYDHS		RV LQNA				EN
Hum_PTP_xi_D1		GITADSSNH		PDNKH		KNRYINIVAYDHS		RVKLAQL				AEK
Hum_PTP_zela_D1		GITADSSNH		PDNKH		KNRYINIVAYDHS		RVKLAQL				AEK
Hum_PTP_gamma_D		NITAEHSNH		PDNKH		KNRYINILAYDHS		RVKLRPL				PGK
Dros_PTP99A_D1		DLPCHEHSQ		HPENKR		KNRYLNITAYDHS		RVHLHPT				PGQ
Hum_LCA_D1		QFTWENSNL		EVNKP		KNRYANVIAVDHS		RVILTSI				DGV
Hum_PTP_mu_D1		SAPWDSAK		DDENRM		KNRYGNIIAYDHS		RVRLQTI				EGD
Hum_PTP_alpha_D1		QATCEAAS		KEENKE		KNRYVNILPYDHS		RVHLTPV				EGV
Hum_PTP_opsilon_D		QCTFELAN		KEENRE		KNRYPNILPNBHS		RVILSQL				DGI
Mouso_CD45_D1		KFPIKDARK		PHNQ		KNRYVDILPYDYN		RVELSEI				NGD
Hum_SH.PTP2		LYSRKEGQ		RQENKN		KNRYKNILPFDHT		RVVLHDG				DPN
Hum_SH.PTP1		LHORLEGQ		RPNKG		KNRYKNILPFDHS		RVILQGR				DSN
Hum_PTP_bola		NQSCDIAL		LPENRG		KNRYNNILPYDAT		RVKLSNV				DDD
Dros_PTP10D		DQPCTFAD		LPENRP		KNRYNNILPYDHS		RVKLPV				DDD
Hum_SAP.1		SQSQMVAS		ASENNA		KNRYRNVL		PDWSRVPLKPI				HEE
Ral_PTP_STEP		FVDPKE		YDIPGLVR		KNRYKTILENPHS		RVRLTSP				DPE
Dros_PTP69A_D1		DRITKNSD		LKENAC		KNRYPDIKAYDQ		TRVKLAVI				NGL
Hum_MEG2		VGTFHCMS		PENLE		KNRYGDPCLDQ		TRVKLTR				SGH
Hum_PTP.PEST		IYPTATGE		KEENVK		KNRYKDILPFDHS		RVKLTLLK				TPS
Hum_PTPH1		GLAITFAK		LQNL		KNRYKDLVLPYD		TRVLLQGN				EDY
Dici_PTP1		PSETSEGD		KKHTS		KNRYTNILPVNHT		RVQLKKI				QDK
Fiss_yeast_pyp1		QWSTVDSL		SNTSYK		KNRYTDIVPYNCT		RVHLKRT				SPS
Fiss_yeast_pyp2		WCCLASSR		STSISR		KNRYTDIVPYDK		TRVRLAVP				KGC
Hum_PTP_xi_D2		GITADSSN		HPDNKH		KNRYINIVAYDHS		RVKLAQL				AEK
Hum_LCA_D2		TSRFISAN		LPENKF		KNRLVNIMPYEL		TRVCLQPI				RGV
Hum_PTP_alpha_D2		NDKMR		TGNL	PANMK	KNRVLQIIPYEF		NRVIIIPVK				RGE
Hum_PTP_opsilon_D2		KENMR		TGNL	PANMK	KARVIQIIPYD		NRVILSMK				RGQ
Hum_PTP_mu_D2		VEDCSIAL		LPNHE		KNRCMDILPPDR		CLPFLITI				DGE
Mouse_CD45_D2		WRTQHI		GNQENKK		KNRNSNVVPYD		NRVPLKHELE				MSKESPESESDDDSD
Dros_PTP69A_D2		SKSCSVGE		NEENNM		KNRSQEIIPYD		NRNRVILTP				PMR
Hum_PTP_zela_D2		QSDYSAAL		KQCNRE		KNRTSSIIIPV		ERSVGISSL				SGE
Hum_PTP_gamma_D2		VECFSAQ		KECNKE		KNRNSVVPSE		RAVGLAPL				PGM
Dros_PTP99A_D2		ETNLMAEO		VEELKNCT		PYLEOYKNIIOF		OPKDIHASAM				KOVNSIKNRGAIFPIEGSRVHLTPKP
Varsinia_PTP		TNDPRYL		QACGGEKI		LNRFRDIOCC		ROTAVRAD				
PTP1Bseq.no.		30	40	50	60							

PTP1B66
Figure 1B

	140	150	160	170	180	190	200
Hum_PTP1B	EKSLKCA	QYWPQKEBEM	IFEDTNLKLTLISEDIKSYTVLELE				NLTQETREI
Hum_TCPTP	EKESVKCA	QYWP DDQEM	LFKETGFSVKLLSEDKSYTVLQLE				NINSGETRTI
Hum_PTP_xi_D1	EKGRRKCD	QYWP ADGSE	EYGN FLVTQKSVQVLAITYTFLRNTKIKKG				SQGRPSGRV
Hum_PTP_zela_D1	EKGRRKCD	QYWP ADGSE	EYGN FLVTQKSVQVLAITYTFLRNTKIKKG				SQGRPSGRV
Hum_PTP_gamma_D	EKGRRKCD	QYWP TENSE	EYGN IIVTLKSTKIHACTYVFSIRNTKVKKGKGNPKGRQNERVV				KKQCNTEKLV
Dros_PTP99A_D1	ERGRKCD	MYWP KDQVE	TYGV IQVLIIEEVMSTYTVLQIKHLKLLK				KSGSSEKREL
Hum_LCA_D1	EKSRVKCD	QYWP ARGTE	TCGL IQVTLDTVELATYTVFALH				KRGVHEIREI
Hum_PTP_mu_D1	EVGRVKCC	KYWP DDTE	IYKD IKVTLIETELLAAYVIFAVE				DMTNRKPORLI
Hum_PTP_alpha_D1	ERKECKCA	QYWP DQGCW	TYGN IRVSVEDTVLVDTYTVFCIQQVG				PDGCKAPRLV
Hum_PTP_opsilon_D	ERKECKCH	QYWP DQGCW	TYGN IRVSVEDCVLVDTYTVFCIQPQL				KKEKATGREV
Mouso_CD45_D1	EGNRNKCA	EYWPMEEGTR	AFKD IIVTINDHKRCPDYIILNVAH				VGQGNTERTV
Hum_SH_PTP2	ERGSKCV	KYWP EYALK	EYGV MRVRNVKESAADHYTLKLSK				LDNGDLIREI
Hum_SH_PTP1	EKGRNKCV	PYWP VGMQR	AYGP YSVTNCGEHDTTEYKLLQVSP				EEQLDAHRLI
Hum_PTP_bola	EKGRVKCD	HYWPA DQDSL	YFGD LILQMLSESVLPWTIFKICG				RGSEQRIL
Dros_PTP10D	EKGREKCD	QYWP DTVPV	FYGD IKVQILNDSHYADWVMFMC				QVEEQKTLV
Hum_SAP.1	EAGRVKCE	HYWPL DSQPC	THGH LRVTLVGEEVMENWTVLLLL				RGTEERGL
Ral_PTP STEP	EMN EKCT	EYWP EEQV	VHDG VEITVQKVIHTEDYRLISLR				VGEEDRRQI
Dros_PTP69A_D1	EYNKAKCA	KYWPKEVFTK	QFGD ILVKAQERKTGDYIELNVSKNAN				NTEERQKRQV
Hum_MEG2	EGRRKCG	QYWPLEKDSRI	RFGF LTVTNLGVENNMHYKKLEIH				FQNESRRL
Hum_PTP. PEST	EMGRKKCE	RYWPLYGEDPI	TFAP FKISCEDEQARTDYFILLLE				NTQTGEEHTV
Hum_PTPH1	ERGRTKCH	QYWP PDVDM	NHGG FHIQCSQSEDCTIAYVSMVLT				LTFEGETRDI
Dici_PTP1	ENCRIKCD	RYWPEQIGGEQFSIYGNENVEFGTSVELVEVIQCREIITRNIR					NANFPSVKV
Fiss_yeast_pyp1	EAGREMCT	AYWPSNGIGDK	QVYGDYCVKQISEENVDSRFLFEIQ				DKPNGPCKXI
Fiss_yeast_pyp2	EAGSEKCS	QYWPDKHACLEGG	LRISVOKYETFEEDLKVHLRL				SQGRPSGRV
Hum_PTP_xi_D2	EKGRRKCD	QYWP ADGSE	EYGN FLVTQKSVQVLAITYTVFLRNTKIKKG				DARDGQSRTI
Hum_LCA_D2	EMGREKCH	QYWP AERSA	RYQY FVVDPMALAYNMPQYILFKVT				NTRNKSROI
Hum_PTP_alpha_D2	ERGQEKCA	QYWP SDGLV	SYGD ITVELKKEBEECESYTVLLVT				ARQEEQVRV
Hum_PTP_opsilon_D2	EREQDKCY	QYWP TEGSV	THGE ITIEIKNDTISEAISIFLVTINQPO				ARPQDGYRMV
Mouse_CD45_D2	PA QLCP	QYWP ENGVH	RHGP IQVEFVSADLEEDIISFRINYA				HSKRKEPRTV
Dros_PTP69A_D2	NGDQEVCA	QYW GEGKQ	TYGD MEVEMKDTNRASAYTLFELR				NCKIDDTLKV
Hum_PTP_zeia_D2	D GPRKCP	RYWA DDEVQ	YDH ILVKVHSESCPYTYFFVT				ATQDDYVLEV
Hum_PTP_gamma_D2	NMAEDEFV	YWPNKDEPNCESFKVTLMAEHLKLSNEEKLIIFILE					ATQDDYVLEV
Dros_PTP99A_D2	SLAEDEFV	YWPRESMNCFAFTVTLISKDRCLCSNEEQIIIFILE					SIODDYELTY
Yarsinia_PTP	D INFA	OFWEDEATPIESDHX	RVKFLNKTKNSDYVSEVIO				REAGOKTISY
PTP1Bseq.no.	ETANOREGMPDYR	OSGT	YGSITVESKMTQOVLGGINMYTLTI				



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Hum_PTP1B	LHFHYTTWPDF	G	VPESPASFLNLFKVR	ES	GSLSP	EHG	PVVVHCSAGIGRSGTFC	270		
Hum_TCPTP	SHFHYTTWPDF	G	VPESPASFLNLFKVR	ES	GSLNPDH	G	PAVTHCSAGIGRSGTFS	260		
Hum_PTP_xi_D1	TQHYTQWPD	M	G	VPEYSLPVLTFVRKA	AYA	KRH	AVG	PVVVHCSAGVGRGTGYI	250	
Hum_PTP_zela_D1	TQHYTQWPD	M	G	VPEYSLPVLTFVRKA	AYA	KRH	AVG	PVVVHCSAGVGRGTGYI	240	
Hum_PTP_gamma_D	IQHYTQWPD	M	G	VPEYALPVLTFVRRS	AA	RMP	ETG	PVLVHCSAGVGRGTGYI	230	
Dros_PTP99A_D1	YQHYTNWPDH	G	TPDHPPLVLFVKKSS	AA	NPA	EAG	EAG	PIVVHCSAGVGRGTGYI	220	
Hum_LCA_D1	RQFQFMAWPDH	G	VPEYPTPILAFLLRR	VKAC	NPL	DAG	DAG	PMVVHCSAGVGRGTGCFI	210	
Hum_PTP_mu_D1	RQFHFTGWPDH	G	VPYHATGLLGFVRQV	KSK	SPP	SAG	SAG	PLVVHCSAGAGRTGCFI	200	
Hum_PTP_alpha_D1	TQFHFTSWPDF	G	VPFTPIGMLKFLKKV	KAC	NPQ	YAG	YAG	AIVVHCSAGVGRGTGFV	190	
Hum_PTP_opsilon_D	SQFHFTSWPDF	G	VPFTPIGMLKFLKKV	KAC	NPQ	HAG	HAG	PIVVHCSAGVGRGTGTFI	180	
Mouso_CD45_D1	THIQFTSWPDH	G	VPEDPHLLKLLRRRV	NAF	SNF	FSG	FSG	PIVVHCSAGIGRTGTFI	170	
Hum_SH_PTP2	WQYHFTWPDH	G	VPSPDGGVLDLFEEV	HHK	QESIM	DAG	DAG	PVVVHCSAGIGRTGTII	160	
Hum_SH_PTP1	WHYQYLSWPDH	G	VPSEPGGVLSFLDQ	INQR	QESLPH	AG	AG	PIIVHCSAGIGRTGTII	150	
Hum_PTP_bola	RHFHYTWPDH	G	VPETTSLSIQFVRTV	RDY	INRSP	GAG	GAG	PTVVHCSAGVGRGTGTFI	140	
Dros_PTP10D	RHFHFTTWPDF	G	VPNPPQTLVRFVRAF	RDR	ICA	EQR	EQR	PIVVHCSAGVGRSGTFI	130	
Hum_SAP.1	RQFHYQAWPDH	G	VPSSPDTLALAFWR	MLRQ	LDQTM	EGG	EGG	PPIVHCSAGVGRGTGLI	120	
Ral_PTP_STEP	KHYWFTSWPDQ	K	TPDRAPPLHLVREVE	EAAQ	QEGPH	CS	CS	PIIVHCSAGIGRTGCFI	110	
Dros_PTP69A_D1	TQHYLTWKDF	M	APHPHGIIFIRQINS	VYSLQ	RG	RG	RG	PILVHCSAGVGRGTGLV	100	
Hum_MEG2	THFQFLSWPDY	G	VPSSAASLIDFLRV	VRNQSLAVSNM	GARSKG	QCPE	PP	PIVVHCSAGIGRTGTFC	90	
Hum_PTP_PEST	YQFHYVNWPDH	G	VPSSFDSILDMLSL	MRKYQ	EH	DV	DV	PICIHCSAGCGRTGAIC	80	
Hum_PTPH1	THLQYVAVPDH	G	IPDDSSDFLEFVNY	VRSLR	VDSE	VDSE	VDSE	PVLVHCSAGIGRTGLV	70	
Dici_PTP1	TQYQYEGWPDH	N	IPDHTQPPRQLLH	SITNRQ	NIIP	SSD	SSD	RNVPIIVHCSAGVGRGTGFC	60	
Fiss_yeast_pyp1	HHYQYPNWSDC	N	SPENVKSMVEFLK	YVNNSH	GSG	GSG	GSG	NTIVHCSAGVGRGTGFI	50	
Fiss_yeast_pyp2	HHFWHTWFD	K	THPDIESITGLIRC	IDKVP	NDG	NDG	NDG	PMFVHCSAGVGRGTGFI	40	
Hum_PTP_xi_D2	TQHYTQWPD	M	G	VPEYSLPVLTFVRKA	AYA	KRH	AVG	PVVVHCSAGVGRGTGYI	30	
Hum_LCA_D2	RQFQFTDWPEQ	G	VPKTGEFIDFIGV	HKT	KEQFG	QDG	QDG	PITVHCSAGVGRGTGVFI	20	
Hum_PTP_alpha_D2	RQFHFGWPEV	G	IPSDGKGMISIIAA	VQKQ	QQQ	SGNH	SGNH	PITVHCSAGAGRTGTFC	10	
Hum_PTP_opsilon_D2	RQFHFGWPEI	G	IPAEKGMDILIAA	VQKQ	QQQ	TGNH	TGNH	PITVHCSAGAGRTGTFI	0	
Hum_PTP_mu_D2	QOQFPLGWPMYRD	TPVSKRSFLKIR	QVQDKWQEE	YNGEG	YNGEG	YNGEG	YNGEG	PTVVHCLNGGGRSGTFC	-10	
Mouse_CD45_D2	YQYQCTTWKGE	E	LPAEPKDLVSMIQ	DLKQKLP	KASPE	GMYK	YH	KHASILVHCRDgSQQTGLFC	-20	
Dros_PTP69A_D2	TQFQYNGWPTVD	GEVPEVC	RGIIELDVQAYNH	YKNNK	NSGC	NSGC	NSGC	RSPLTVHCSLgTDRSSIFV	-30	
Hum_PTP_zeia_D2	RHFQCPKWP	N	PDSPISKTFFELIS	VIKEEA	NR	DG	DG	PMIVHDEHGVTAGTFC	-40	
Hum_PTP_gamma_D2	RHFQCPKWP	N	PDAPISSTFELIN	VIKEEA	TR	DG	DG	PTIVHDEYGAVSAGMLC	-50	
Dros_PTP99A_D2	KMLHCPSPWPM	SNPNSIYDFIVD	VHERCNDY	RNG	RNG	RNG	RNG	PIVIVDRYGGAOACTFC	-60	
Yarsinia_PTP	PVVHVGNWPOTAV	SSVETKALASL	VDOTAETKRN	MYESK	GSSA	VADD	SKLR	PVIHCRAGVGRTAOLI	-70	
PTP1Bseq.no.	180	190	200	210	220	230	240	250	260	270

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Figure 1D

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Hum_PTP1B	280	LADTCLLMDDR	290	KOPSSVDI	300	KKVLLMRKFRMG	310	LIQTADQLRFSYLAVIEGAKFIMGD	320	330	340
Hum_TCPTP		LVDTCCLVMEKG		DD		INI		KQVLLNMRKFRMG			
Hum_PTP_xi_D1		VLDMLQIQIHE		GT		VNI		FGFLKHIRSQRNY			
Hum_PTP_zela_D1		VLDMLQIQIHE		GT		VNI		FGFLKHIRSQRNY			
Hum_PTP_gamma_D		VLDMLQIQIKDK		ST		VNV		LGFLKHIRTQRNY			
Dros_PTP99A_D1		VLDAMLKQIQK		NI		VNV		FGFLRHIAQRNF			
Hum_LCA_D1		VIDAMLERMKHE		KT		VDI		YGHVTCMRQRNY			
Hum_PTP_mu_D1		VIDIMLDMARE		GV		VDI		YNCVRELRSRVN			
Hum_PTP_alpha_D1		VIDAMLDMMHTE		RK		VDV		YGFVSRIARQCQ			
Hum_PTP_opsilon_D		VIDAMMAMHAE		QK		VDV		FEFVSRIARQRPQ			
Mouso_CD45_D1		GIDAMLEGLEAE		GK		VDV		YGVVVKLRQRCL			
Hum_SH.PTP2		VIDILIDIIREK		GVD		DDI		PKTIQMVRQRSG			
Hum_SH.PTP1		VIDMLMENISTK		GL		DDI		QKTIQMVRQRSG			
Hum_PTP_bola		ALDRILQQLDSK		DS		VDI		YGAVHDLRLHRVH			
Dros_PTP10D		TLDRILQQLINTS		DY		VDI		FGIVYAMRKEVRW			
Hum_SAP.1		ALDVLLRQLQSE		GL		LGP		FSFVRKMRRESRPL			
Ral_PTP_STEP		ATSIQCQQLRRE		GV		VDI		LKTTQQLRQDRGG			
Dros_PTP69A_D1		ALDSLIIQQLLEE		DS		VSI		YNTVCDLRHQRNF			
Hum_MEG2		SLDICLAQLEEL		GT		LVN		FQTVSRMRTQRAF			
Hum_PTP.PEST		AIDYTWNLLKAG		KIPEEFNV		FNLIQEMRTQRHS		AVQTKQYELVHRAIAQFIEKQLQLY			
Hum_PTPH1		TMETAMCLTERN		LP		IYP		LDIVRKMRDQRAM			
Dici_PTP1		TAVIMMKKLDHYFKQLDYNRIDENL		FSIVLKLREQRPG		MVQQLQYLCYKTYLDEIYHRLNC		MVQQLQYLCYKTYLDEIYHRLNC			
Fiss_yeast_pyp1		VLDTILRFPESKLSGFNPSVADSSDVFLVDHIRKQRMK		MVQTFQFKYVD		LIDS					
Fiss_yeast_pyp2		AVDOILQVPKNLLPK		TJNLEDSKFIENCNSLSORMK		MVONFEOKFLXD		VVDXL			
Hum_PTP_xi_D2		VLDMLQIQIHE		GT		VNI		FGFLKHIRSQRNY			
Hum_LCA_D2		TLSTVLERMRYE		GV		VDM		FQTVKTLRTQRP			
Hum_PTP_alpha_D2		ALSTVLERVKAE		GI		LDV		FQTVKSLRLQRP			
Hum_PTP_opsilon_D2		ALSNILERVKAE		GL		LDV		FQAVKSLRLQRP			
Hum_PTP_mu_D2		ALSIVCCEMLRHQ		RT		VDV		FHAVKTLRNNKPN			
Mouse_CD45_D2		ALFNLLESAETE		DV		VDV		FQVVKSLRKARPG			
Dros_PTP69A_D2		AMCITLVQHLE		KC		VDI		CATTIKLRSQRTG			
Hum_PTP_zela_D2		ALTTLMHQLEKE		NS		VDV		YQVAKMINLMRPG			
Hum_PTP_gamma_D2		ALTTLSQQLENE		NA		VDV		FQVAKMINLMRPG			
Dros_PTP99A_D2		ALSSLAIEMEYC		ST		ANV		YQYAKLYHNKRP			
Yarsinia_PTP		GAMCMNDNRNSO		LSV		EDMVSOMRVORNG		MVQKDEQLDVLK			
PTP1Bseq.no.											

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Figure 1E

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Vmax and Km of 37kDa-PTP1B Mutants Toward RCML

Enzyme	Vmax (nmol/min/mg)	Km (nM)	Kcat (min ⁻¹)
wild type	60200	102	2244
Tyr 46 → S	4120	1700	154
→ L	4160	1700	155
Glu 115 → A	5700	45	212
→ D	5900	20	220
Lys 116 → A	68600	150	2557
Lys 120 → A	19000	80	708
Asp 181 → A	0.61	≤126	0.023
→ E	97	10	3.6
His 214 → A	700	20	26
Cys 215 → S	0.026		0.00097
Arg 221 → K	11	80	0.41
→ M	3.3	1060	0.12
Gln 262 → A	720	9	27

Figure 2